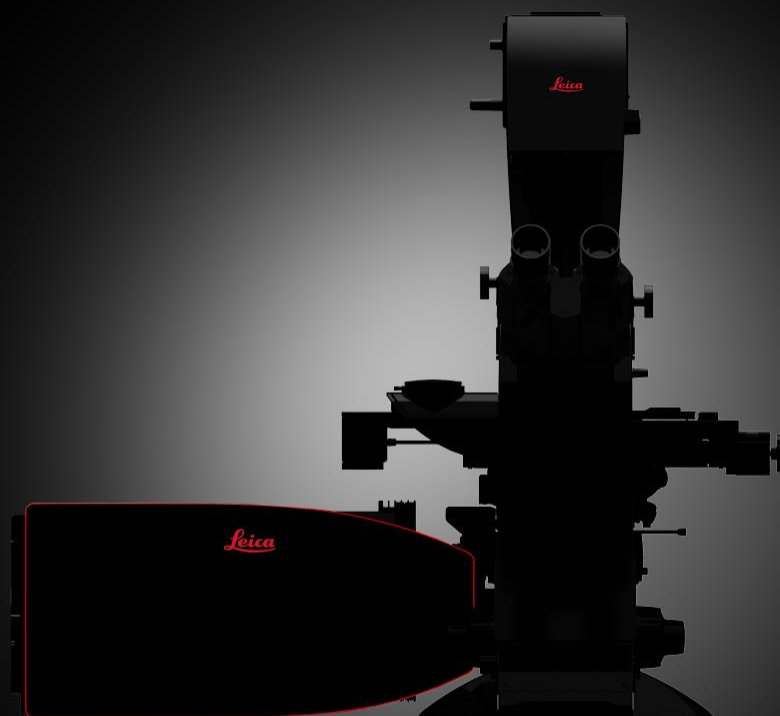




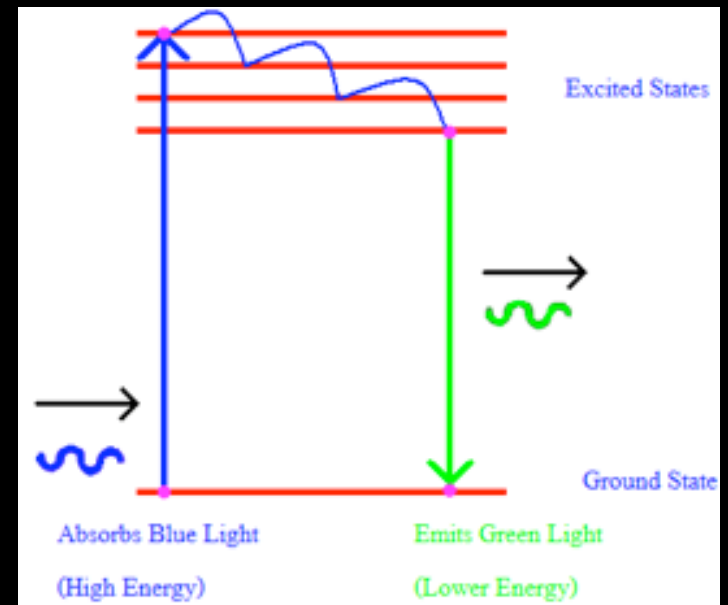
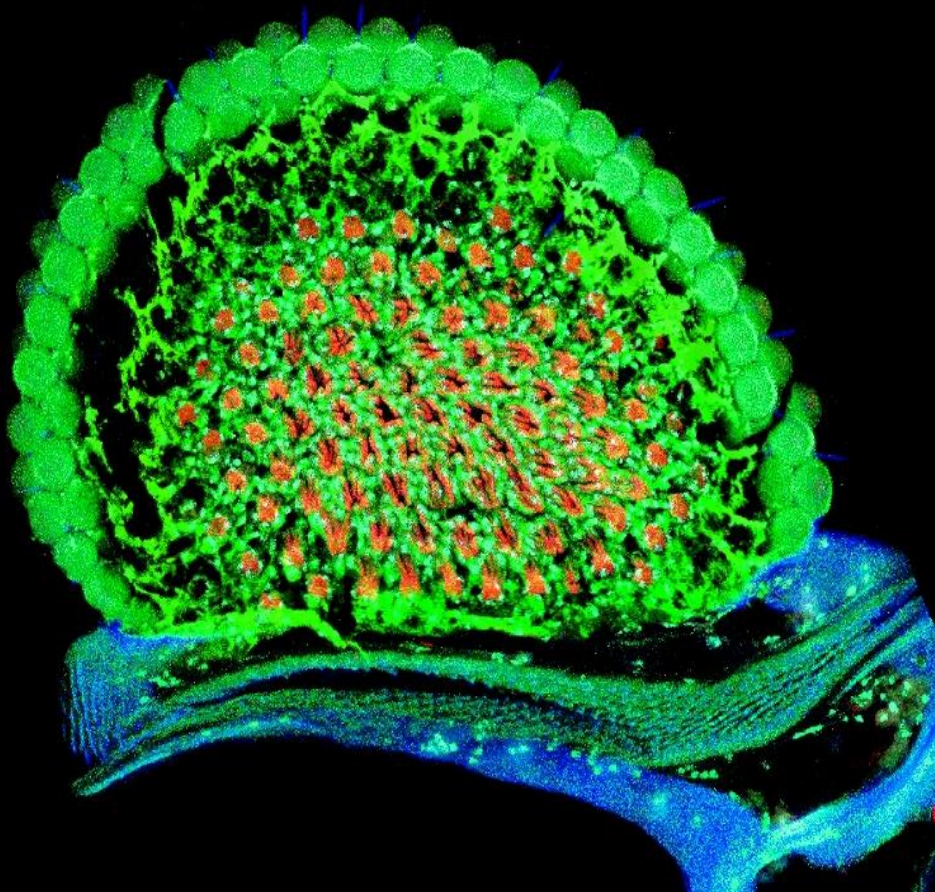
STELLARIS

Leica New Confocal Platform



劉思嫻
美嘉儀器股份有限公司
www.major.com.tw

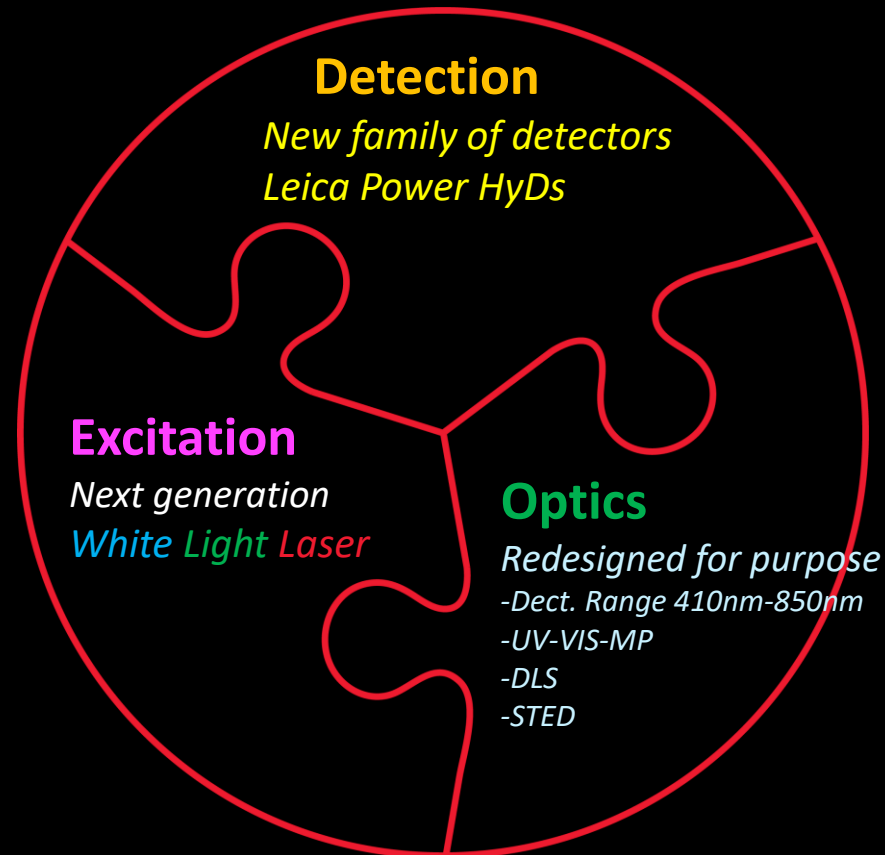
Confocal Image -- Fluorescence



What is Power of Confocal Microscope?

Three key points:

More Sensitivities, More Flexibilities, More Modularizations



The Second Key Innovation: The next-generation White Light Lasers

Traditional Confocal



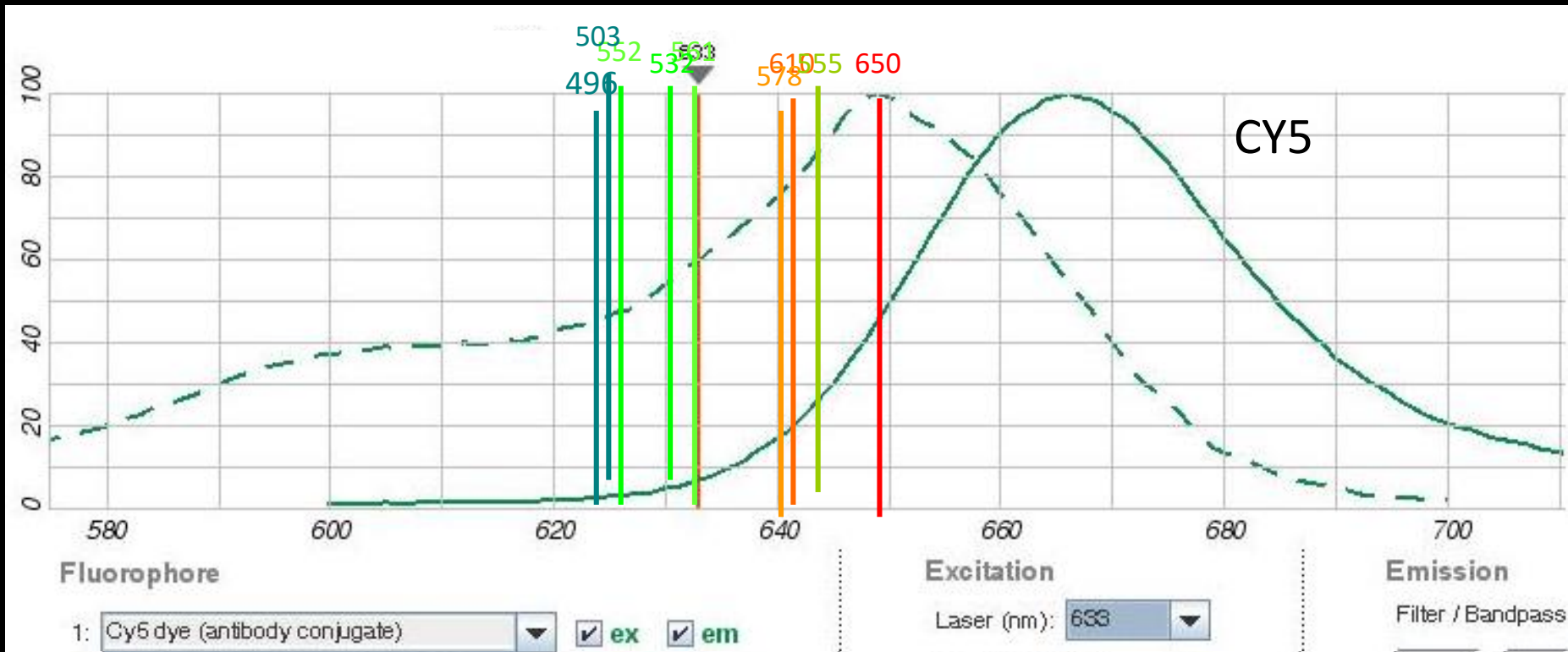
STELLARIS 8



- > Experience complete spectral freedom with excitation perfectly matched to the fluorochrome
- > Less complexity, more flexibility: a single laser to do the work of many. Up to 8 single excitation lines from 440 nm to 790 nm can be used simultaneously
- > New optics design: detection range from 410nm to 850nm

Excitation wavelength

458nm, 476nm, 488nm, 496nm, 514nm, 543nm, 561nm, 594nm, 633nm



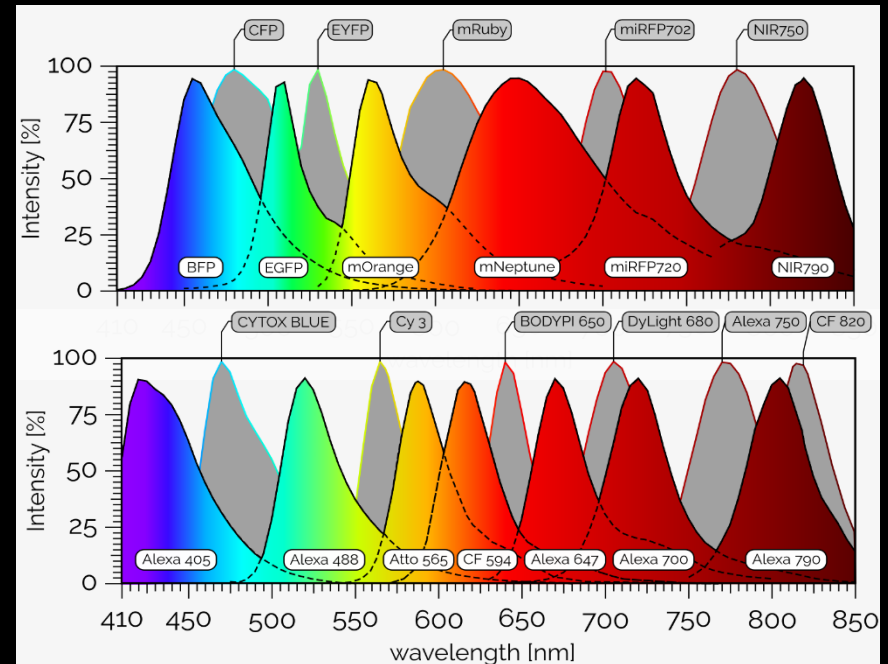
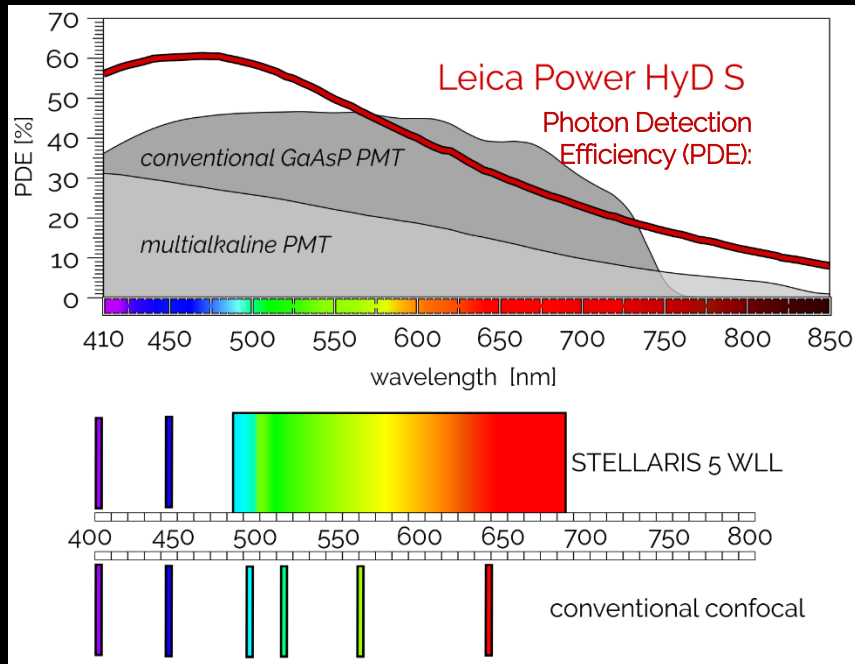
LEICA STELLARIS Confocal Microscope

Excitation wavelength

440nm - 790nm, 351 ex. Lines, no limitation

Enhanced Spectral Freedom: STELLARIS

The Power HyD S Is The New All-rounder Detector For Confocal Applications



- No more PMT or GaAsP detectors
- Detection range: 410nm-----850nm
(normal confocal: 400nm-750nm)

The Red Extended Benefits Of Our Next Generation WLLs

- > Excite each fluorophore optimally at its excitation peak
- > Enhance multiplexing capabilities by adding up to 3 more fluorophores in the NIR range

Some >685 nm excitable dyes:

ATTO 740

ATTO 700

CF680

CellBrite NIR750

Alexa 750

CellBrite NIR680

CF700

CF750

MitoView720

CellBrite NIR770

BioTracker NIR750

Alexa 680

Alexa 700

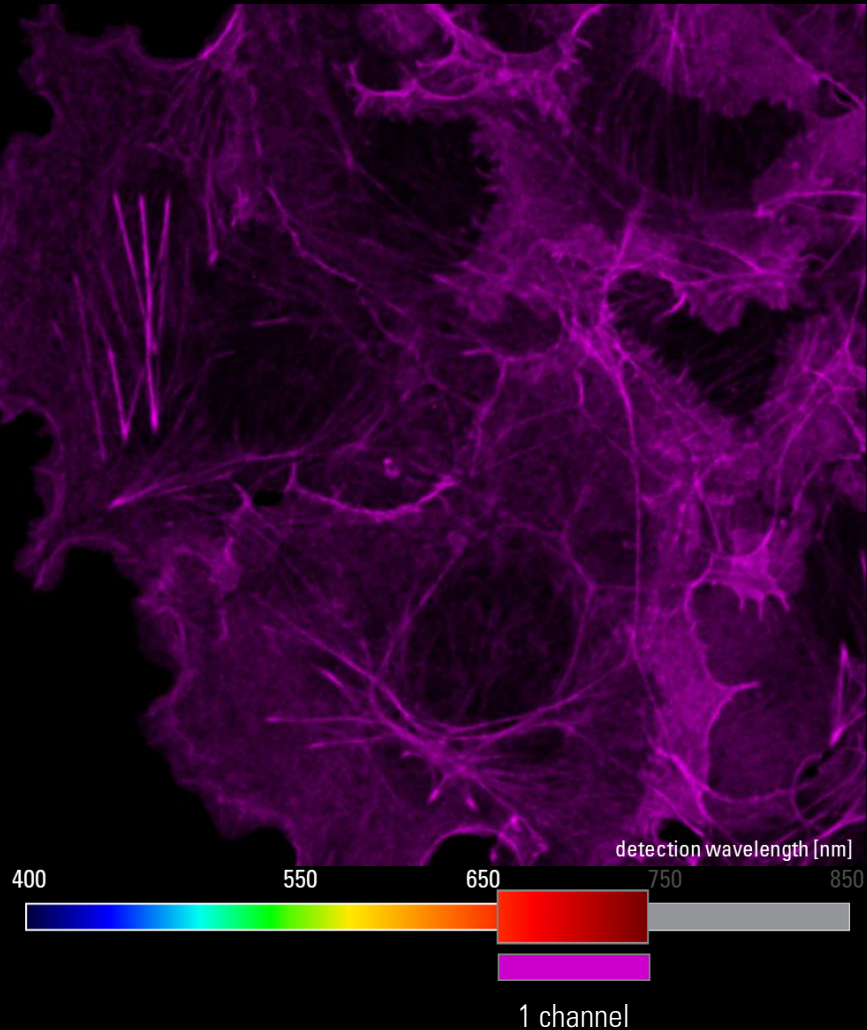
ATTO 680

ATTO 725

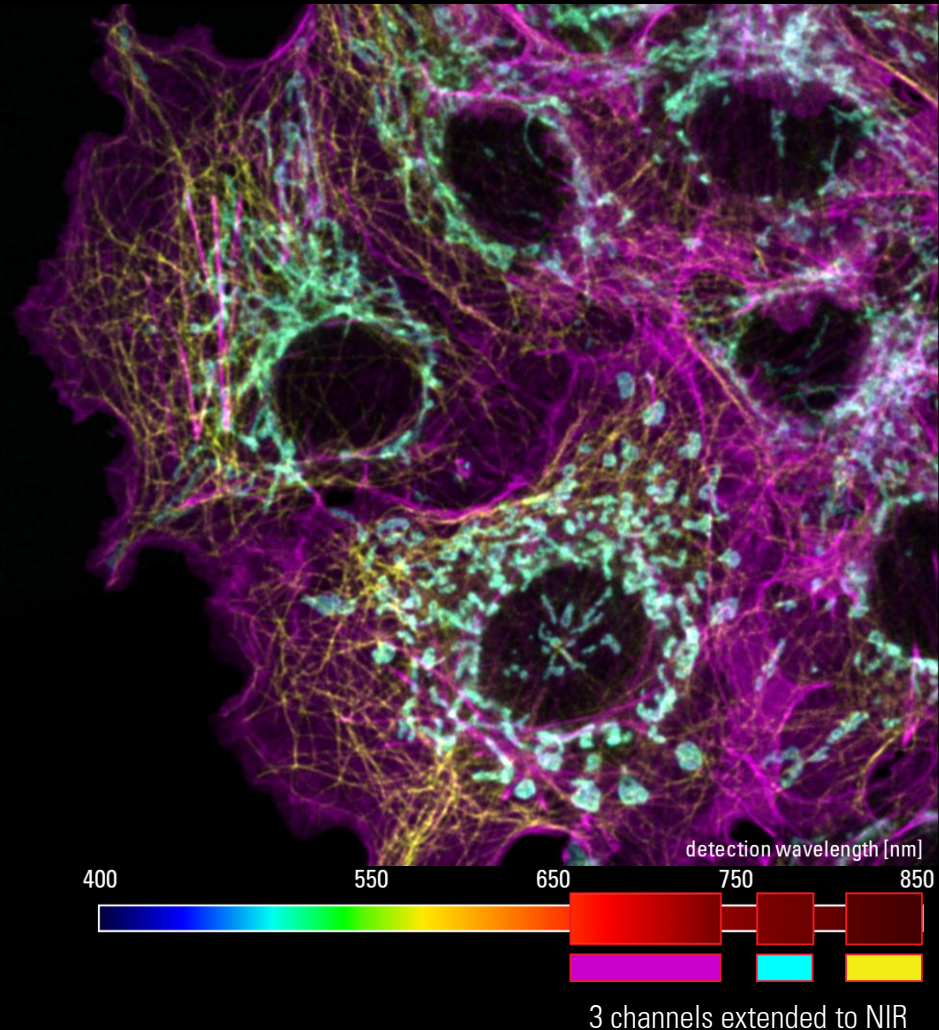
CellBrite NIR700

STELLARIS Delivers Expanded Multicolor Flexibility

Traditional Confocal



STELLARIS

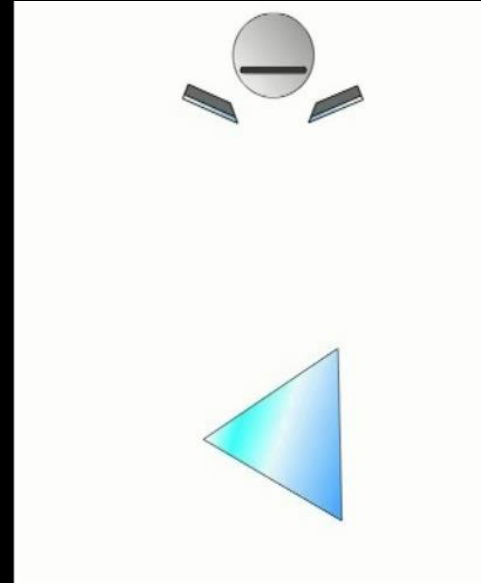
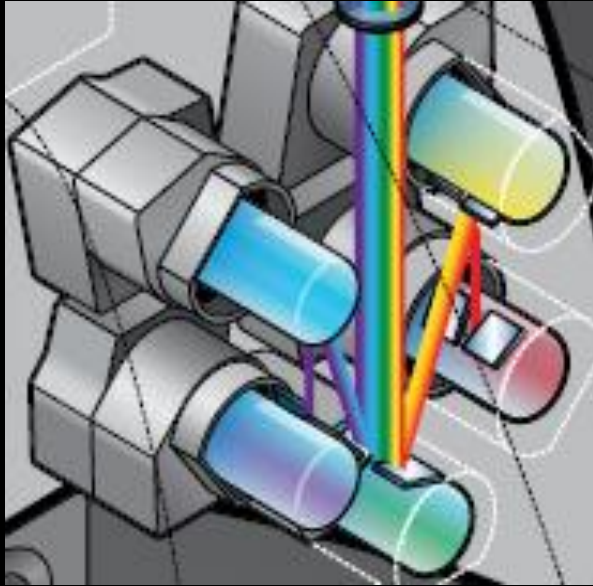


COS7 cells. Actin (magenta, SiR-Actin 657-740 nm), Mitochondria (cyan, AF750 760-790 nm), Microtubules (yellow, AF790 810-850 nm)
Sample Courtesy: Jana Döhner, Urs Ziegler, University of Zurich

Leica



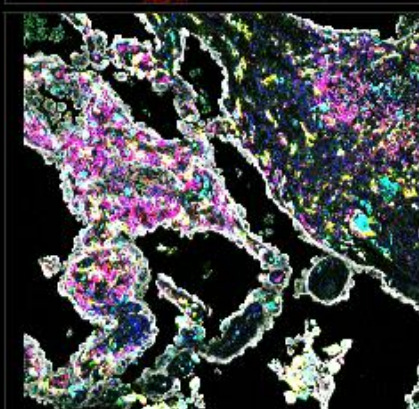
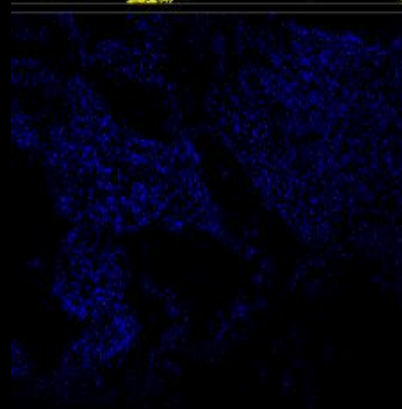
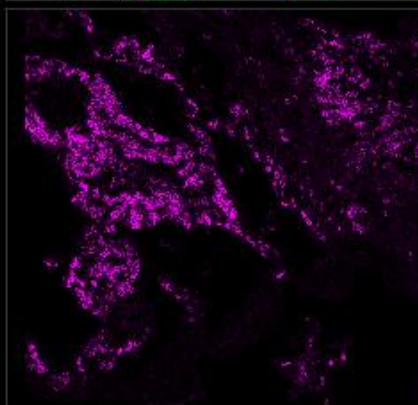
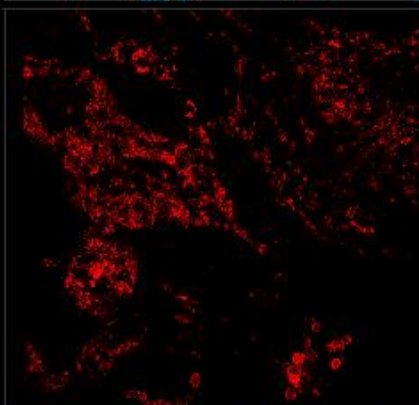
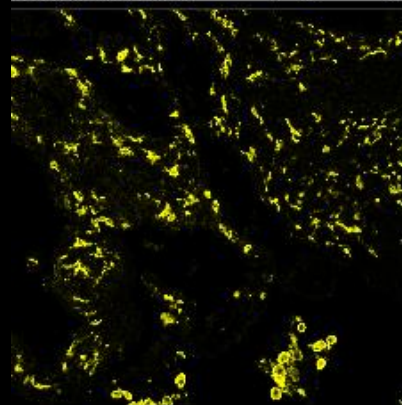
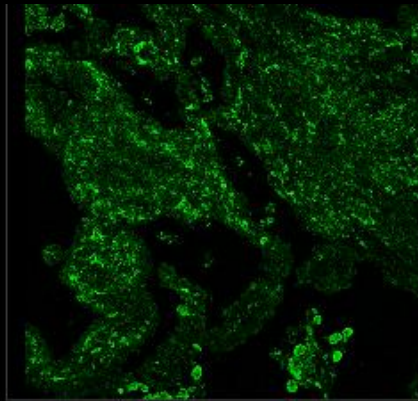
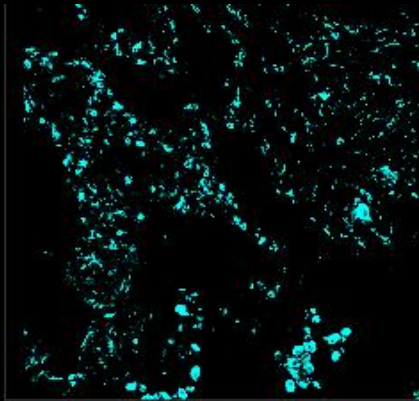
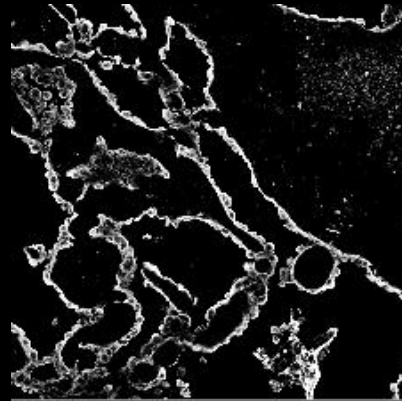
Spectral Imaging Detector -- Fit For Purpose



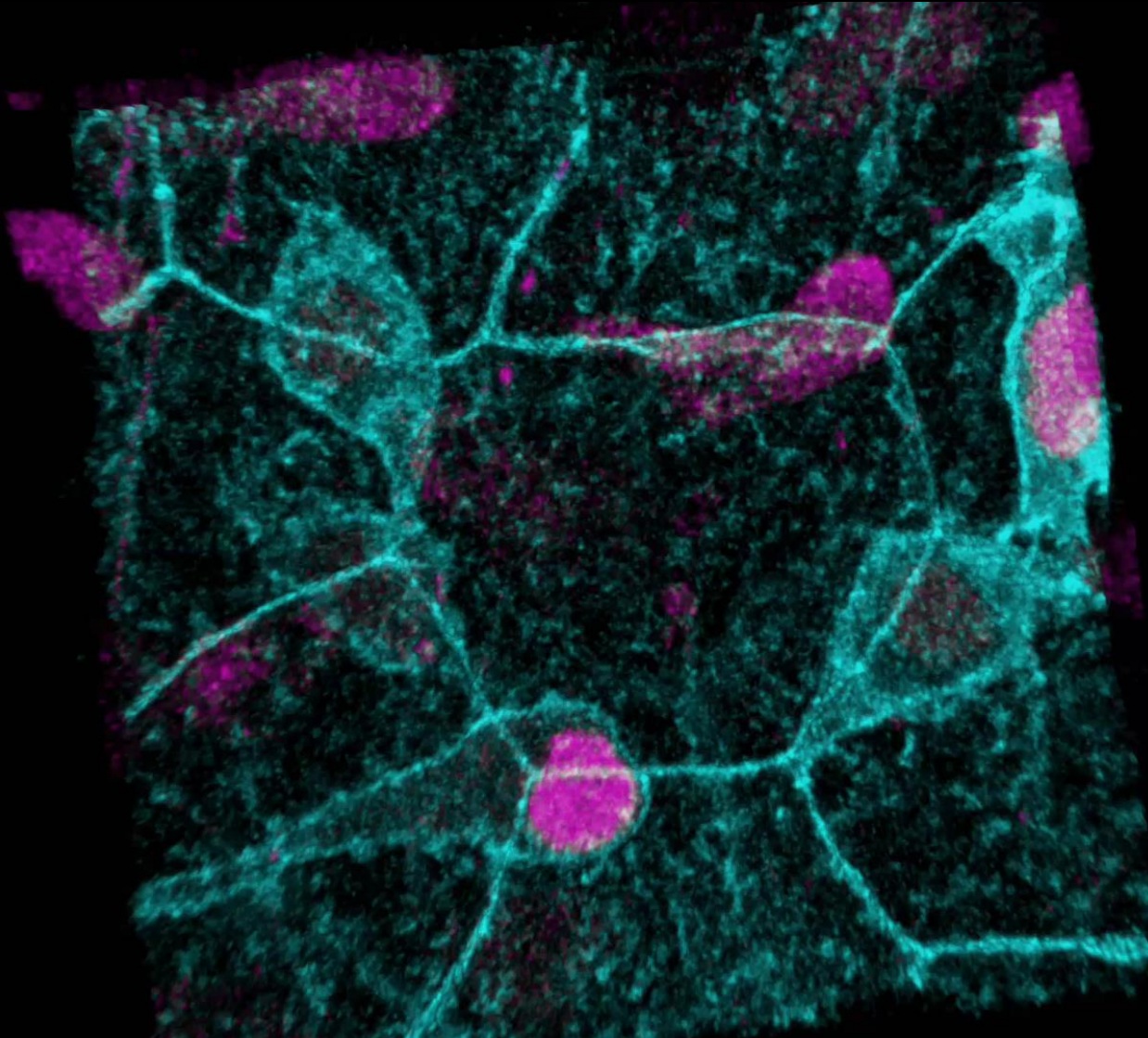
Why White Light Laser System?

OPAL 7 Kit
Opal多重螢光組織染色試劑

Opal Fluorophores	Wavelength	
	Excitation	Emission
Opal 520	494 nm	525 nm
Opal 540	523 nm	536 nm
Opal 570	550 nm	570 nm
Opal 620	588 nm	616 nm
Opal 650	627 nm	650 nm
Opal 690	676 nm	694 nm
Spectral DAPI	405 nm	461 nm



Gentle Live-Cell Imaging

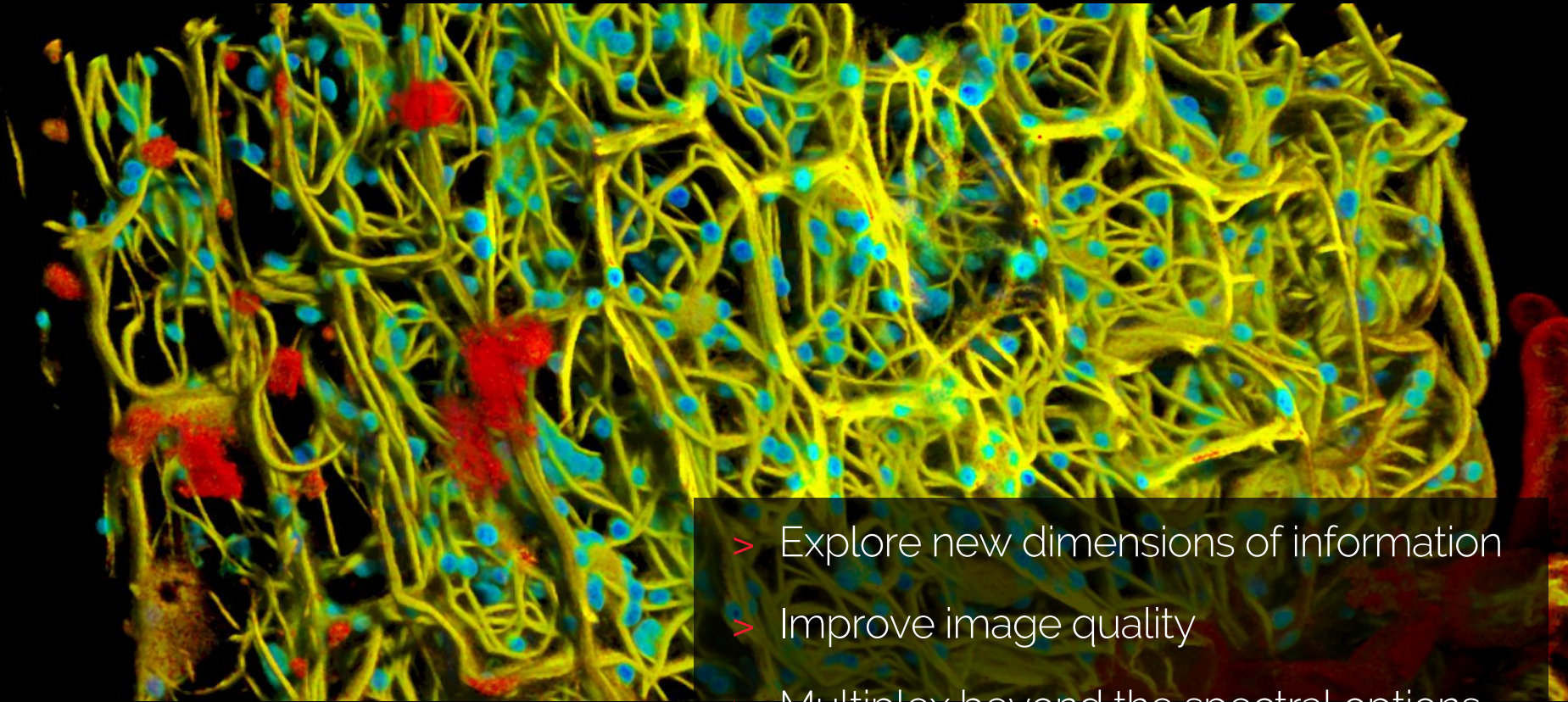


- Perform imaging for longer periods, since both excitation as well as detection are optimally tuned
- Preserve sample integrity with efficient signal acquisition, at the lowest needed power

Zebrafish posterior lateral line primordium migration. Cyan: Membranes, GFP, Magenta: Nuclei, tdTomato
Sample Courtesy: Jonas Hartmann, Gilmour Group, EMBL Heidelberg.

POTENTIAL

DISCOVER MORE



- > Explore new dimensions of information
- > Improve image quality
- > Multiplex beyond the spectral options

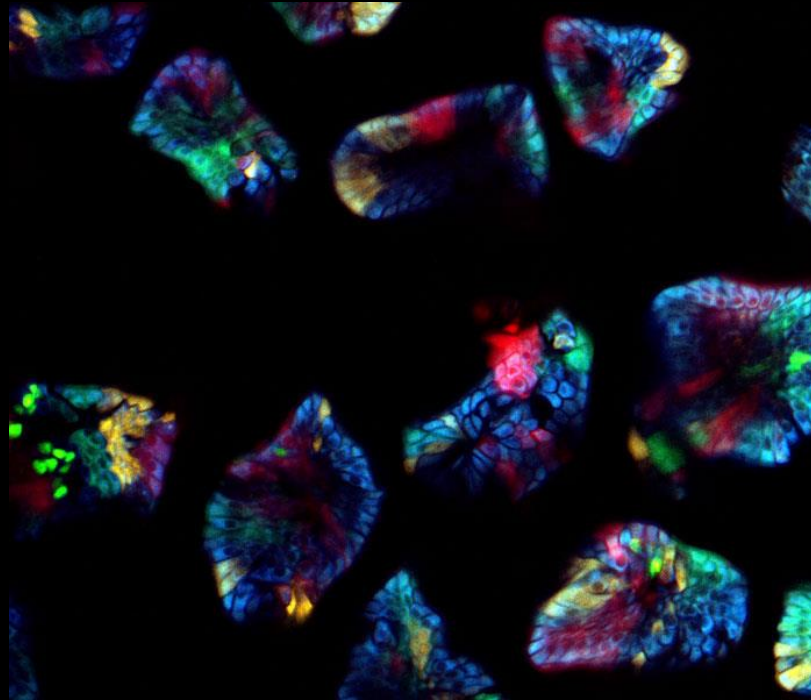
TauSense™

- > Set of tools based on fluorescence lifetime



Leica STELLARIS首度將時脈頻譜
技術 TauSense technology 整合於
共軛焦顯微平台上

Fluorescence imaging focus on spectral contrast....



Confetti Mouse Small Intestine. CFP, GFP, YFP and RFP. Acquired with SP8 DIVE
Sample courtesy of Jacco van Rheenen,
University of Utrecht, the Netherlands

... fluorescence contains much more information

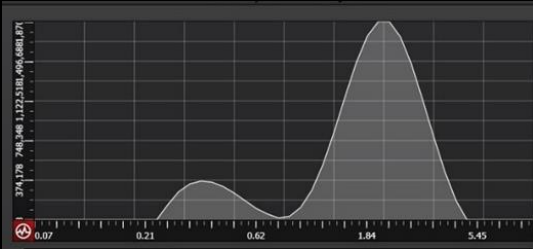
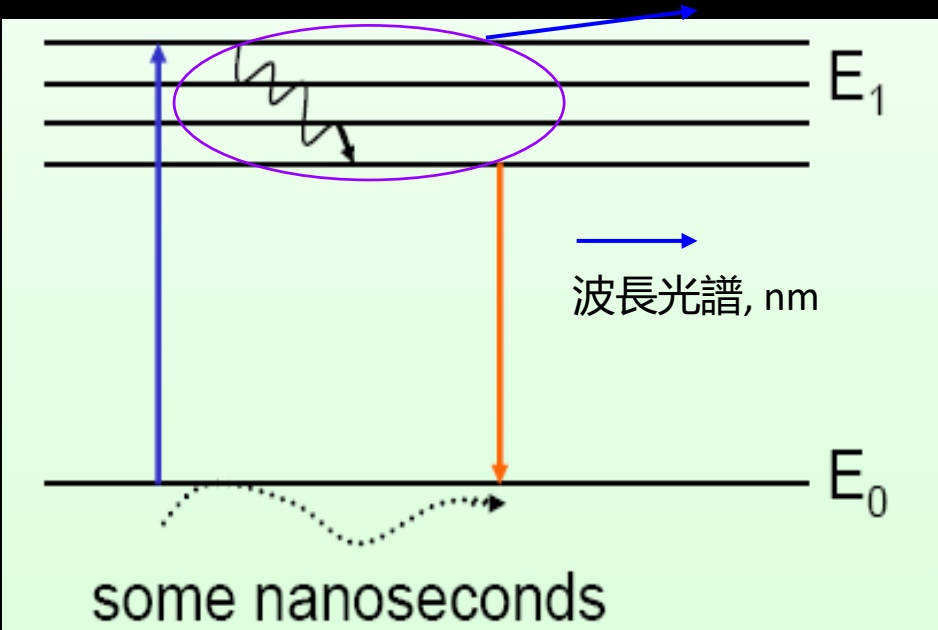
時脈頻譜(TauSense)技術的應用概念來自螢光生命週期影像技術

Fluorescence Lifetime Imaging Microscopy (FLIM)

Fluorescence lifetime

Average time that molecules stay in their excited state

生命週期 (時脈頻譜, ns)



Fluorophore	Ex. Max.	(X) Em. Max.
AF 488	494nm	519nm
GFP	498nm	516nm

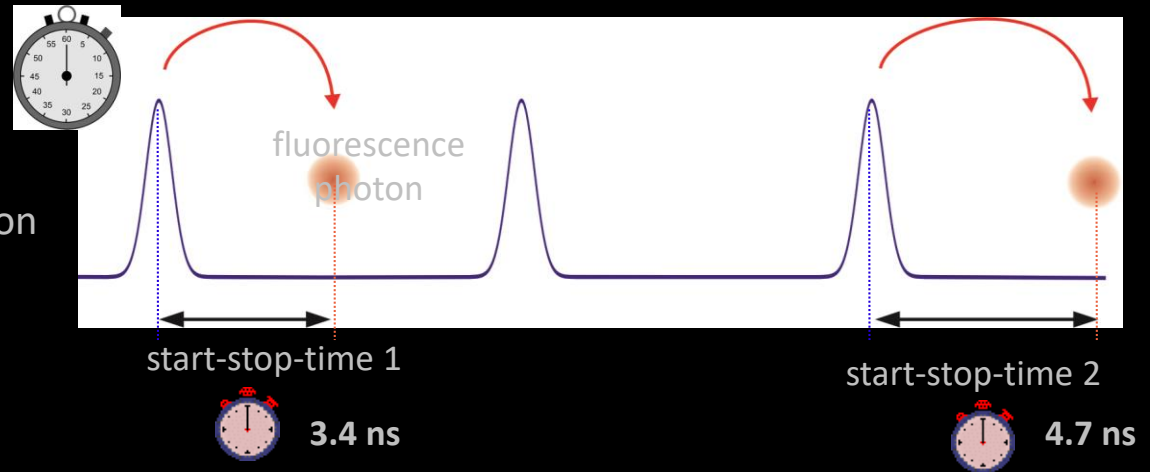
Fluorophore	Ex. Max.	Em. Max.
Fluo3 w. Ca2+	490nm	520nm
Fluo3 w/o Ca2+	490nm	520nm

How to Measure the Time?

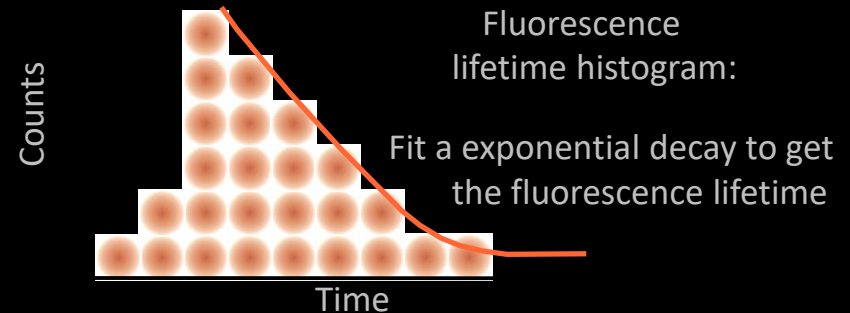
In principle with a stop watch:

1. Start the clock with a laser pulse.
2. Stop the clock with the first photon that arrives at the detector.
3. Reset the clock and wait for next start signal.

It is a statistical process!

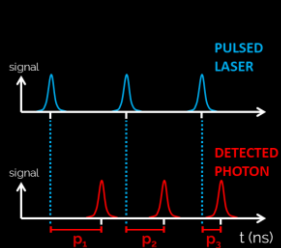


- Repeat this time measurement very often and count “how many photons have arrived after what time”
- Sort the photons within a histogram into time bins according to their arrival times



The Technology Behind TauSense

- > Fluorescence Intensity (N_{photons})
- > Photon Arrival Time (ns)



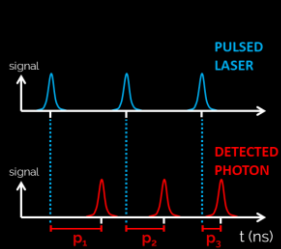
- 1) FPGA
- 2) Pixel-by-pixel
- 3) On the fly

- > Fluorescence Intensity (N_{photons})
- > **Average** Photon Arrival Times (**AAT**, ns)



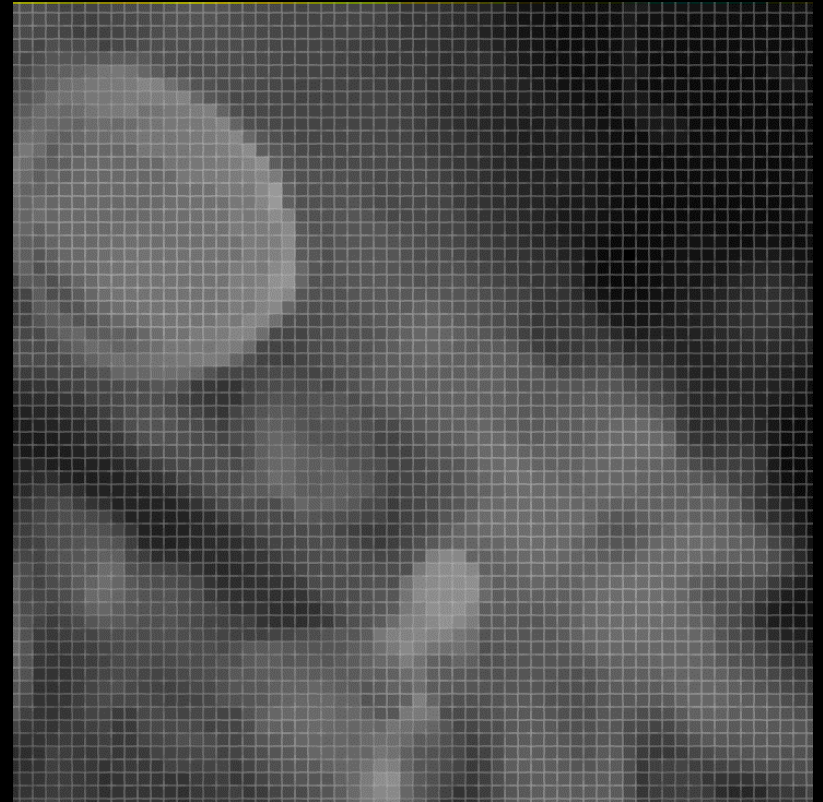
The Technology Behind TauSense

- > Fluorescence Intensity (N_{photons})
- > Photon Arrival Time (ns)



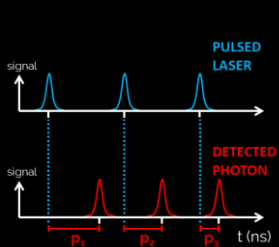
- 1) FPGA
- 2) Pixel-by-pixel
- 3) On the fly

- > Fluorescence Intensity (N_{photons})
- > **Average** Photon Arrival Times (**AAT**, ns)



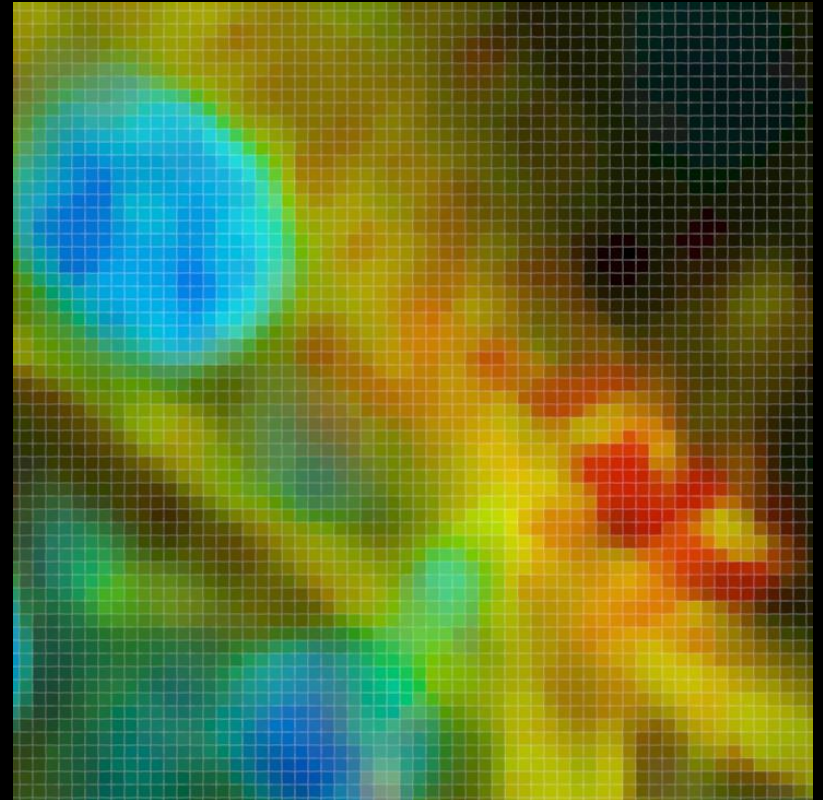
The Technology Behind TauSense

- > Fluorescence Intensity (N_{photons})
- > Photon Arrival Time (ns)



- 1) FPGA
- 2) Pixel-by-pixel
- 3) On the fly

- > Fluorescence Intensity (N_{photons})
- > **Average** Photon Arrival Times (**AAT**, ns)



Alexa Dyes

Alexa Fluor Dye *	Ex (nm)	Em (nm)	τ (ns) ‡
Alexa Fluor 488	496	519	4.1 §
Alexa Fluor 532	532	553	2.5
Alexa Fluor 546	556	573	4.1
Alexa Fluor 555	555	565	0.3
Alexa Fluor 568	578	603	3.6 §
Alexa Fluor 594	590	617	3.9 §
Alexa Fluor 647	650	665	1.0
Alexa Fluor 660	663	690	1.2 **
Alexa Fluor 680	679	702	1.2
Alexa Fluor 700	702	723	1.0
Alexa Fluor 750	749	775	0.7

Measurements were made on free succinimidyl ester derivatives in aqueous solutions. † For Alexa Fluor 488, Alexa Fluor 532, Alexa Fluor 546, Alexa Fluor 555, Alexa Fluor 568, Alexa Fluor 594 and Alexa Fluor 647 dyes, QY measurements were made in PBS (50 mM potassium phosphate, 150 mM NaCl, pH 7.2) at 22°C relative to fluorescein in 0.01 M NaOH (QY = 0.92). For Alexa Fluor 660, Alexa Fluor 680, Alexa Fluor 700 and Alexa Fluor 750 dyes, QY measurements were made in PBS (50 mM potassium phosphate, 150 mM NaCl, pH 7.2) at 22°C relative to Alexa Fluor 647 succinimidyl ester in PBS (QY = 0.33). ‡ Except for the footnoted values, lifetime measurements were made in water at 22°C, data provided by ISS Inc. (Champaign, IL). § Lifetime measurements were provided by the SPEX Fluorescence Group, Horiba Jobin Yvon Inc. ** Lifetime measurement was made in pH 7.5 buffer at 20°C by Pierre-Alain Muller, Max Planck Institute for Biophysical Chemistry, Göttingen.



Atto Dyes

Dyes	Ex (nm)	Em (nm)	τ (ns)
Atto 465	453	508	2.2
Atto 488	501	523	3.2
Atto 495	495	527	2.4
Atto 514	511	533	3.0
Atto 520	516	538	3.8
Atto 532	532	553	3.8
Atto Rho6G	535	560	4.1
Atto 550	554	576	3.2
Atto 565	563	592	3.4
Atto Rho3B	565	592	1.5
Atto Rho11	571	595	4.0
Atto Rho12	576	601	4.0
Atto Thio12	579	609	2.0
Atto Rho101	586	610	4.2

Dyes	Ex(nm)	Em(nm)	τ (ns)
Atto 590	594	624	3.7
Atto 594	601	627	3.5
Atto Rho13	600	625	3.9
Atto 610	615	634	3.3
Atto 620	619	643	2.9
Atto Rho14	625	646	3.7
Atto 633	629	657	3.2
Atto 647	645	669	2.3
Atto 647N	644	669	3.4
Atto 655	663	684	1.9
Atto Oxa12	663	684	1.8
Atto 665	663	684	2.9
Atto 680	680	700	1.8
Atto 700	700	719	1.5
Atto 725	729	752	0.5
Atto 740	740	764	0.6



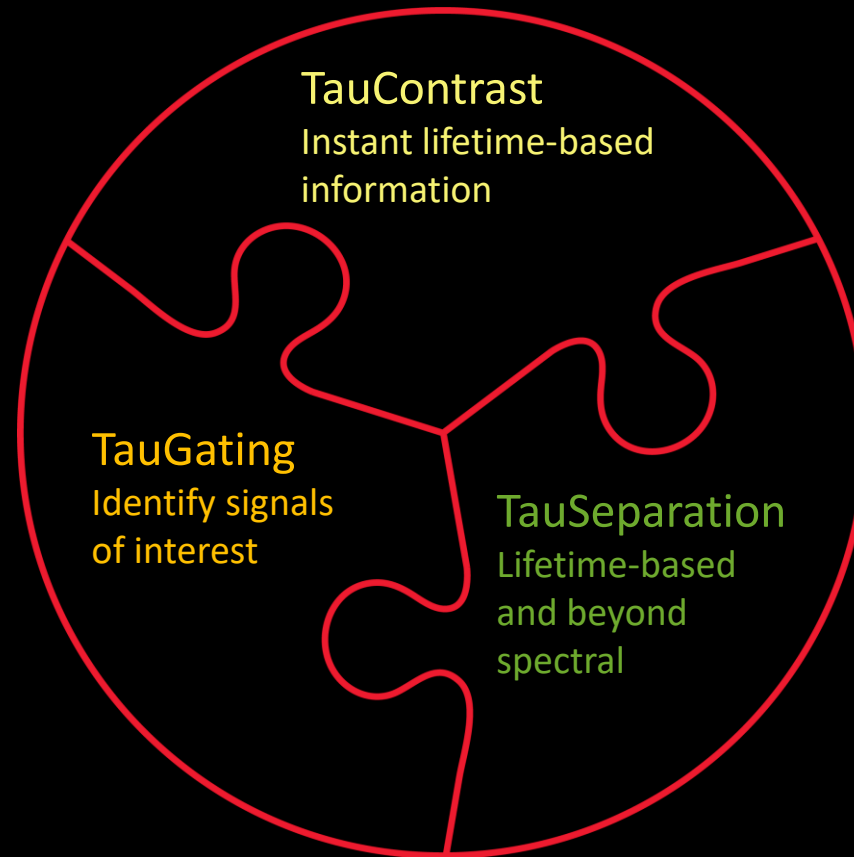
Dye	Ex (nm)	Em (nm)	τ (ns)
Cy3	548	562	0.3
Cy3.5	581	507	2.6
Cy5	646	664	1.0
Cy5.5	675	695	1.0

FITC	494	518	4.1
Oregon Green 488	493	520	4.1
Oregon Green 500	402	522	2.18
Rhodamine 6G	525	555	4.08
Rhodamin B	562	583	1.68
Texas Red	589	615	4.2
TOTO-1	514	533	2.2

Fluorescent Protein

Fluorescent Protein	Ex (nm)	Em (nm)	τ (ns)
ECFP	434	477	3.0
EGFP	488	507	2.6
EYFP	513	527	3.1
mRuby	558	605	2.6
mScarlet	569	594	3.9
mCherry	587	610	1.4
mKate2	588	633	2.5

STELLARIS: Ready to Discover with TauSense (時脈頻譜技術)



ALL Leica STELLARIS series confocal with WLL offer TauSense technology (lifetime-based information) to discover more



Explore A New Dimension Of Information

Traditional Confocal

STELLARIS

TauSense Tool:
TauContrast

Leica

Explore New Dimensions Of Information



Root-hypocotyl-junction of
Arabidopsis thaliana (Era et al.
Plant Cell Physiol., 2009).
Chlorophyll, Life-Act Venus, IProp.
Sample courtesy: Dr. Krebs, COS,
University of Heidelberg.

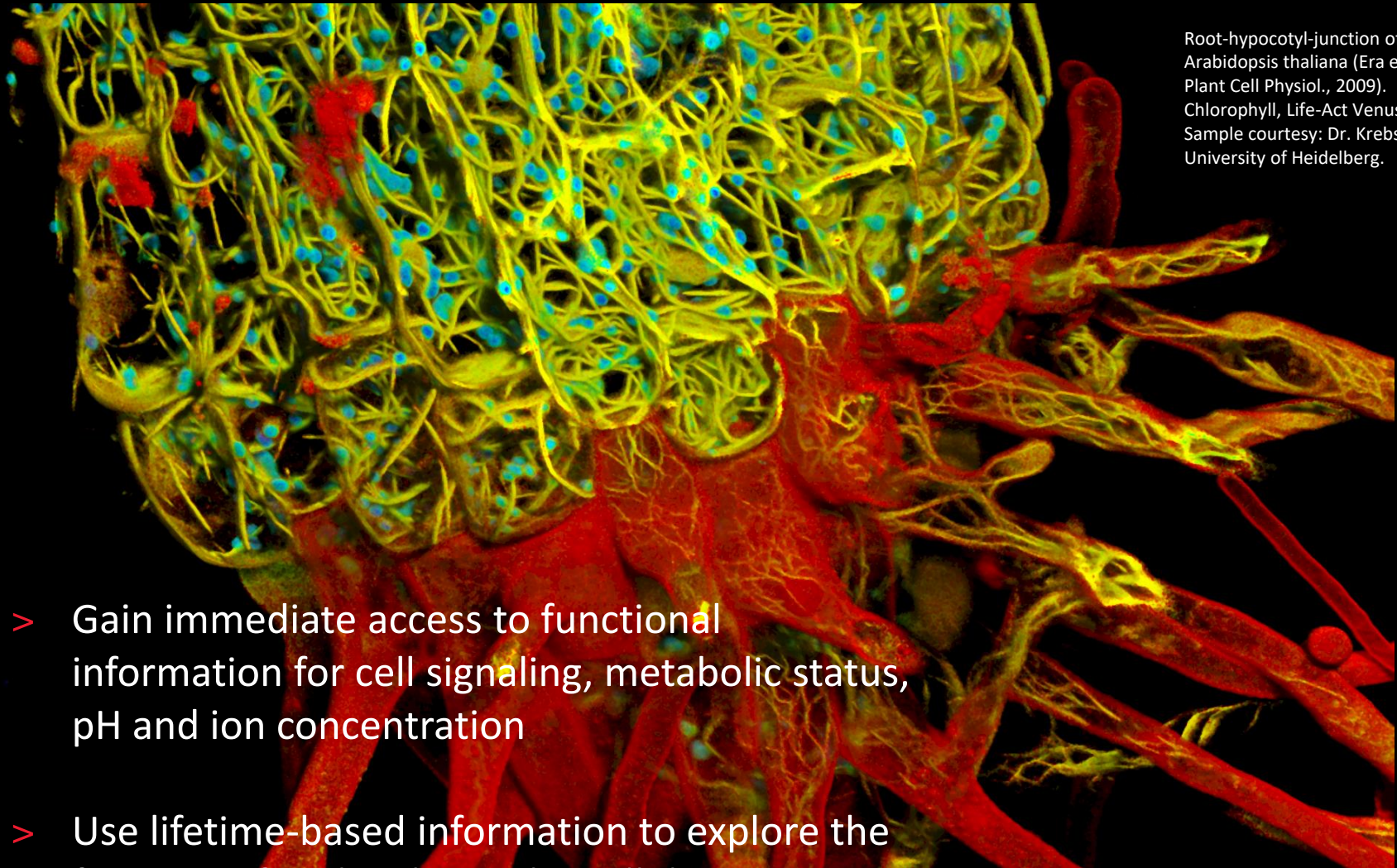
... go beyond today's limits

... see the unseen

... acquire more accurate and reliable data to prove your hypothesis

Leica

Explore New Dimensions Of Information - TauContrast



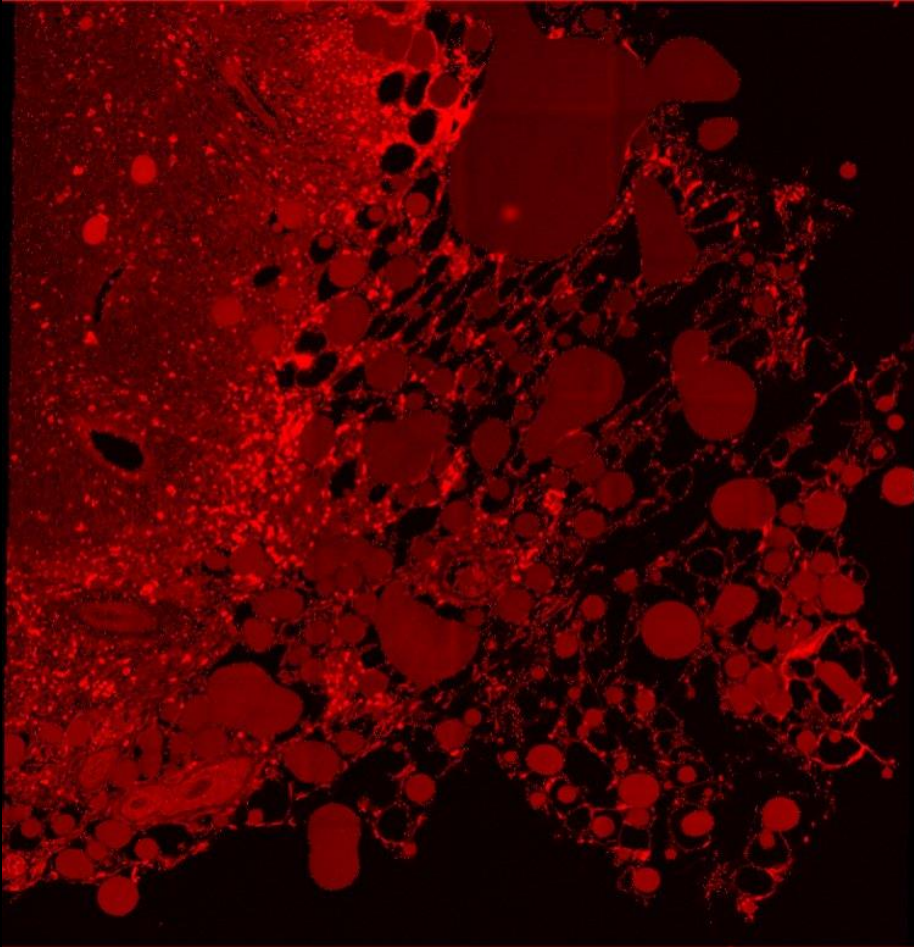
Root-hypocotyl-junction of
Arabidopsis thaliana (Era et al.
Plant Cell Physiol., 2009).
Chlorophyll, Life-Act Venus, IProp.
Sample courtesy: Dr. Krebs, COS,
University of Heidelberg.

- > Gain immediate access to functional information for cell signaling, metabolic status, pH and ion concentration
- > Use lifetime-based information to explore the function of molecules in the cellular context

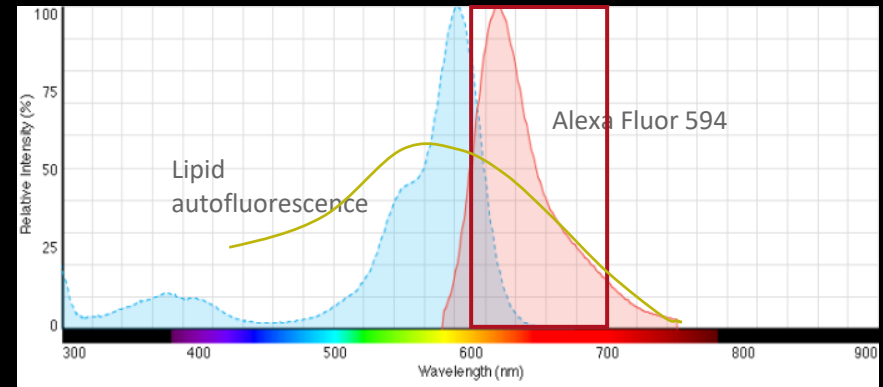
TO GET CLOSER TO THE TRUTH

Leica

TauContrast – autofluorescence- T cell



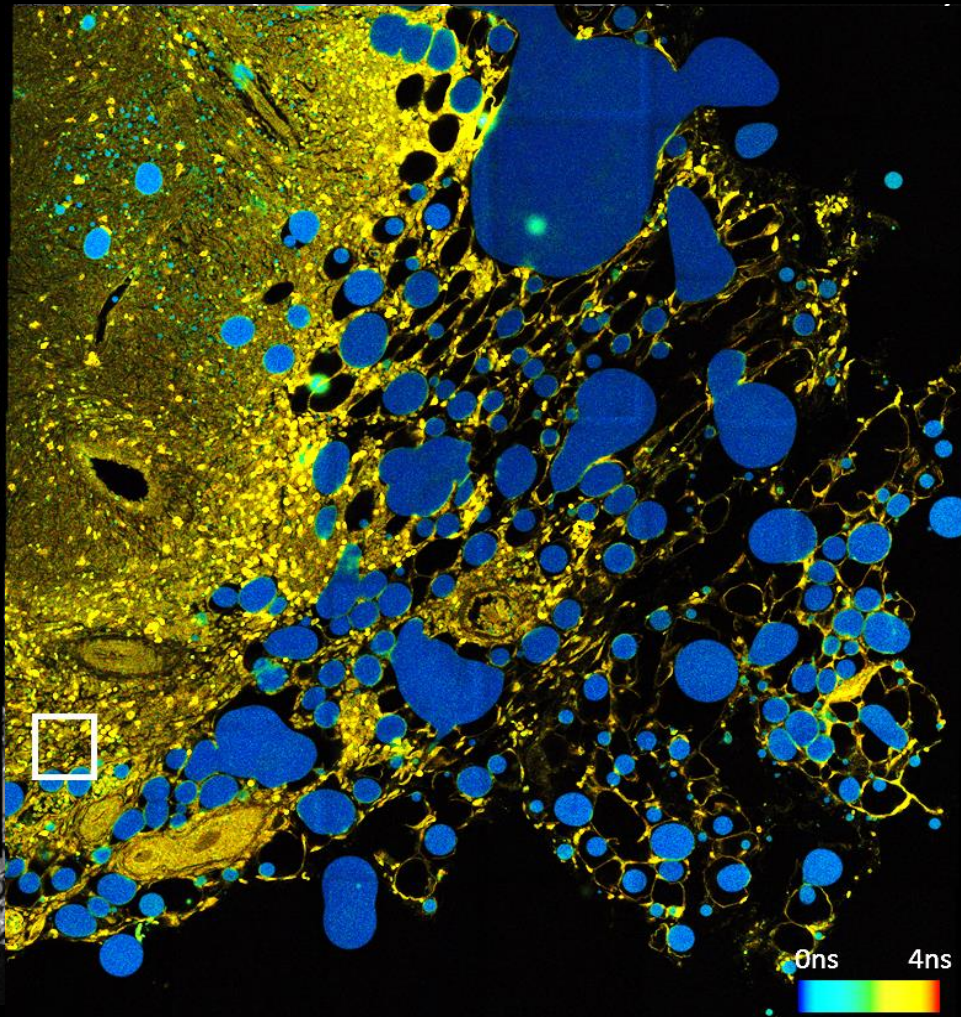
Excitation : 588nm, detection : 615-705nm



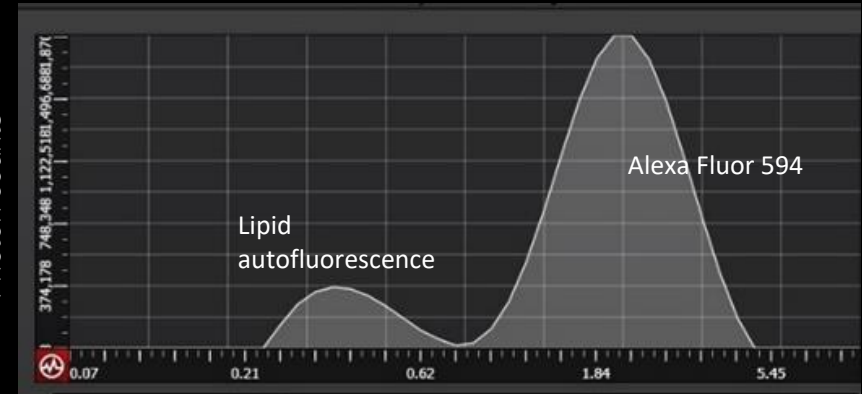
Subcutaneous BRAF600E mouse melanoma with surrounding fat tissue. Cytotoxic T cells stained with anti-CD8a Alexa Fluor 594. Tissue shows high autofluorescence which interferes with Alexa Fluor 594 signal. Fluorescence lifetime information (fast FLIM) enables to distinguish CD8a+ T cells (longer lifetime, yellow) from the autofluorescence of the lipid droplets (short lifetime in blue) and of other cells (green in Fast FLIM images)

Courtesy of Dr. Jan Boettcher, Institute of Molecular Immunology, TU Munich

TauContrast – autofluorescence- T cell



Excitation : 588nm, detection : 615-705nm

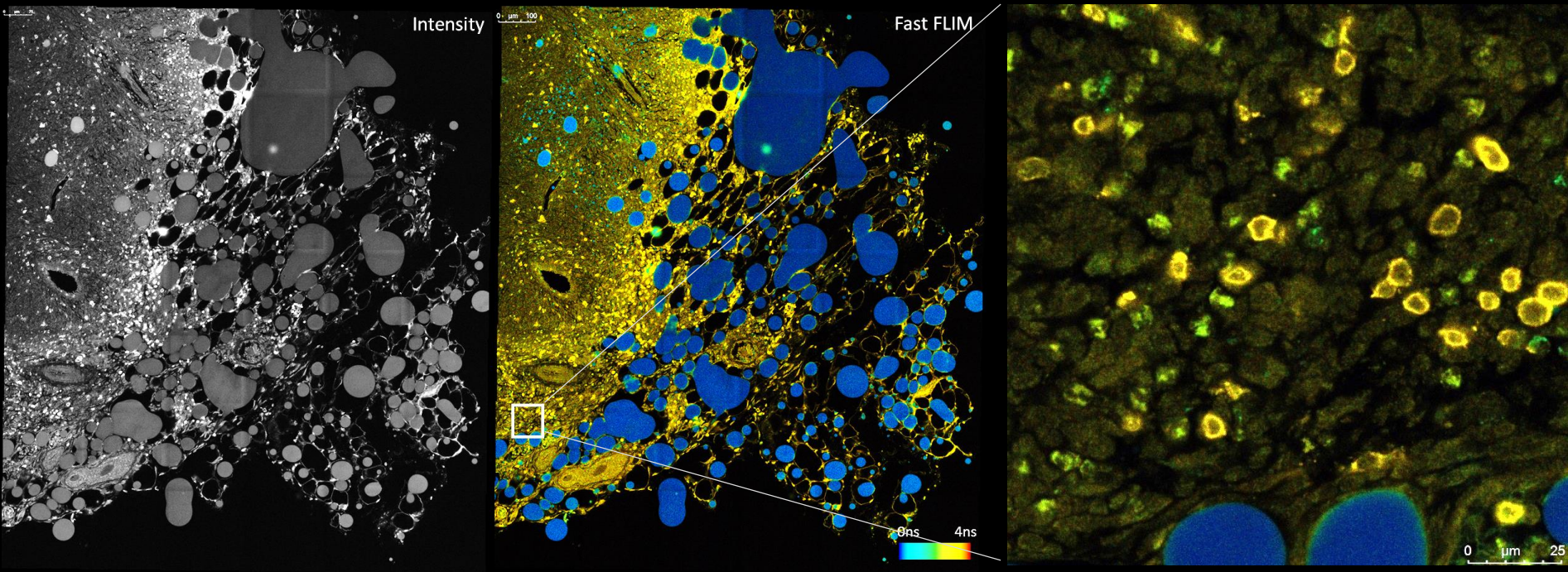


Average arrival time, ns

SUBCUTANEOUS BRAFV600E MOUSE MELANOMA WITH SURROUNDING FAT TISSUE. CYTOTOXIC T CELLS STAINED WITH ANTI-CD8A ALEXA FLUOR 594. TISSUE SHOWS HIGH AUTOFLUORESCENCE WHICH INTERFERES WITH ALEXA FLUOR 594 SIGNAL. FLUORESCENCE LIFETIME INFORMATION (FAST FLIM) ENABLES TO DISTINGUISH CD8A+ T CELLS (LONGER LIFETIME, YELLOW) FROM THE AUTOFLUORESCENCE OF THE LIPID DROPLETS (SHORT LIFETIME IN BLUE) AND OF OTHER CELLS (GREEN IN FAST FLIM IMAGES)

Courtesy of Dr. Jan Boettcher, Institute of Molecular Immunology, TU Munich

TauContrast – autofluorescence- T cell

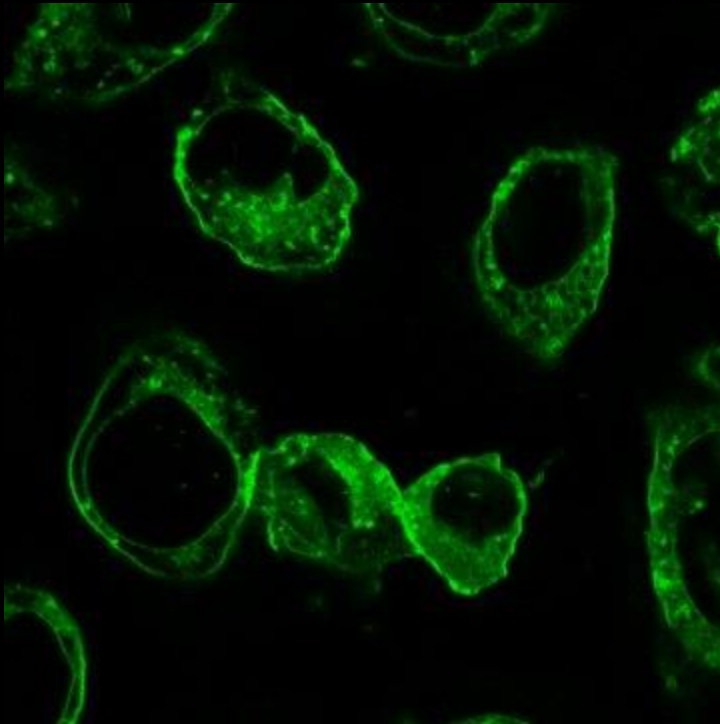


Subcutaneous BRAF600E mouse melanoma with surrounding fat tissue. Cytotoxic T cells stained with anti-CD8a Alexa Fluor 594. Tissue shows high autofluorescence which interferes with Alexa Fluor 594 signal. Fluorescence lifetime information (fast FLIM) enables to distinguish CD8a+ T cells (longer lifetime, yellow) from the autofluorescence of the lipid droplets (short lifetime in blue) and of other cells (green in Fast FLIM images)

Courtesy of Dr. Jan Boettcher, Institute of Molecular Immunology, TU Munich

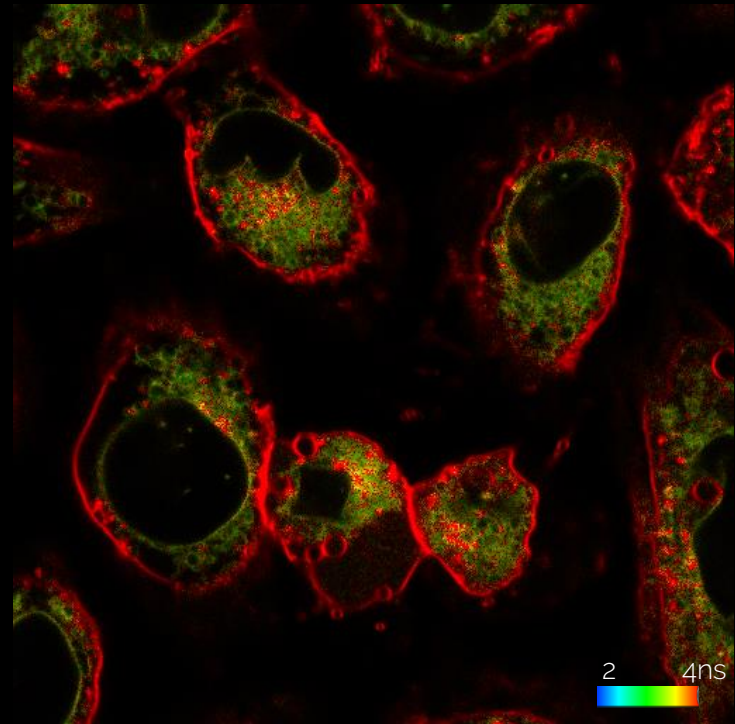
Membrane tension visualization with FlipperTR[®]

Excitation : 488nm, detection : 575-625nm
Intensity



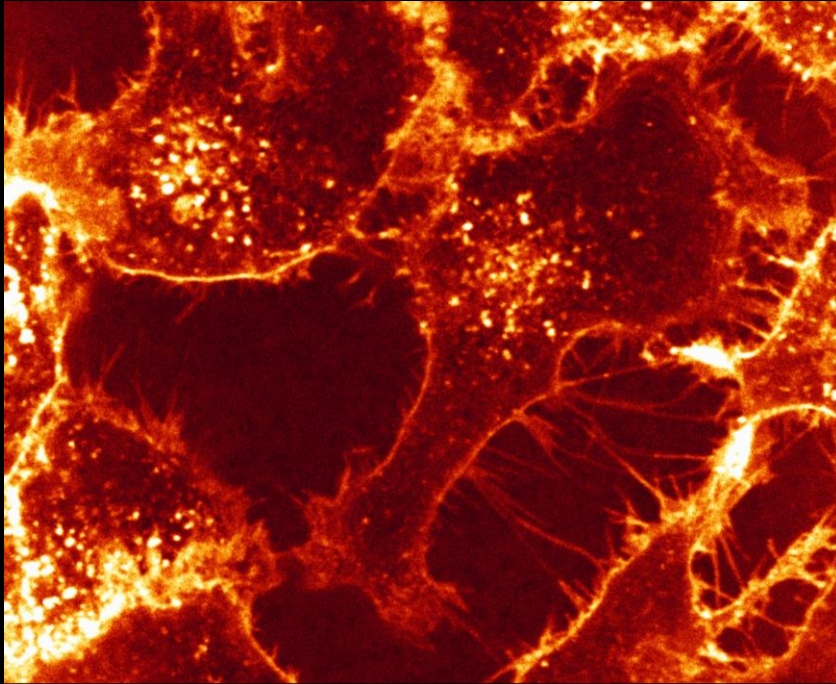
U2OS cells labeled with Flipper TR,

Tau Contrast



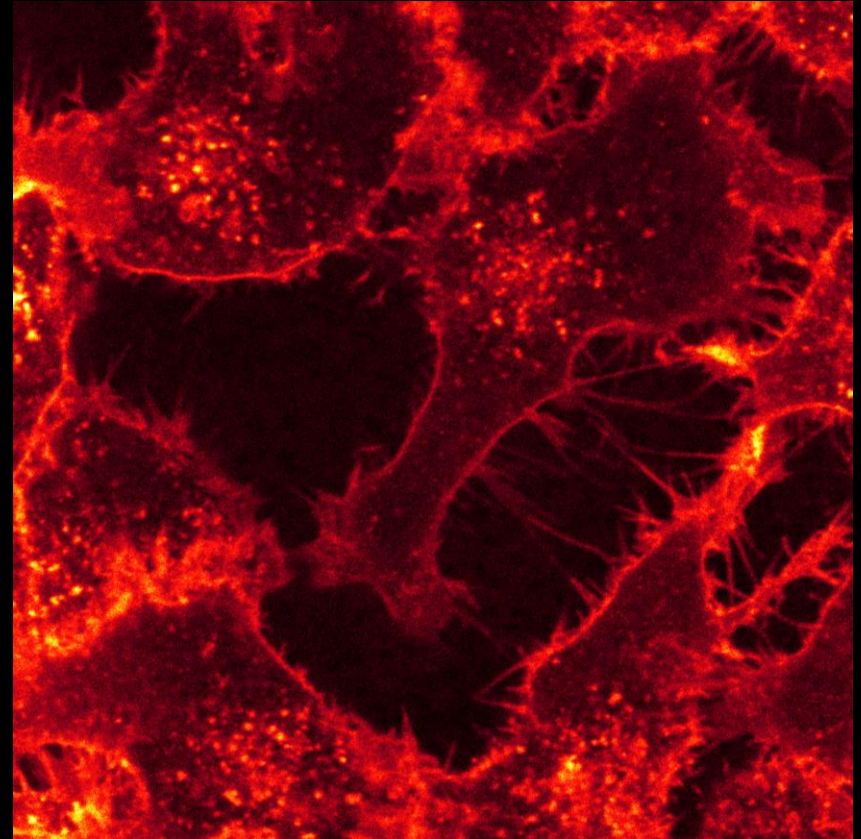
Improve Image Quality

Traditional Confocal

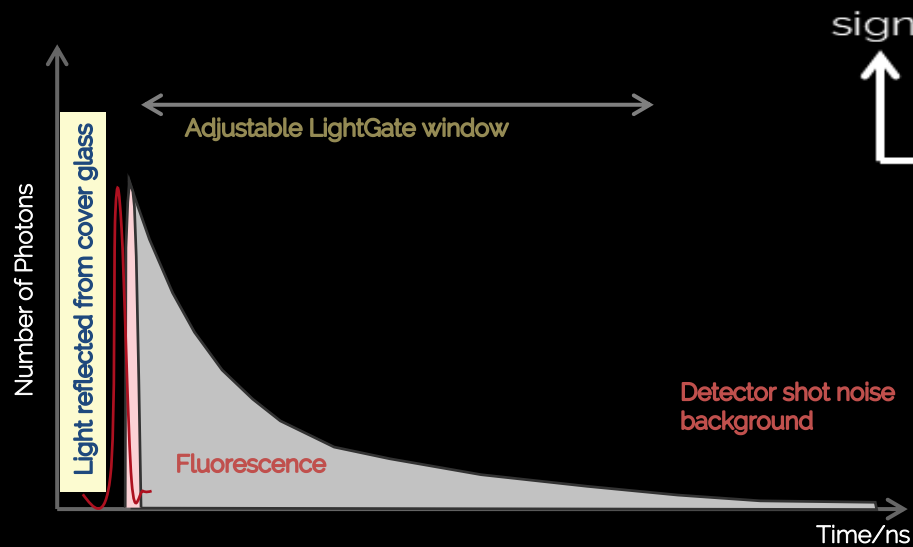


TauSense Tool:
TauGating

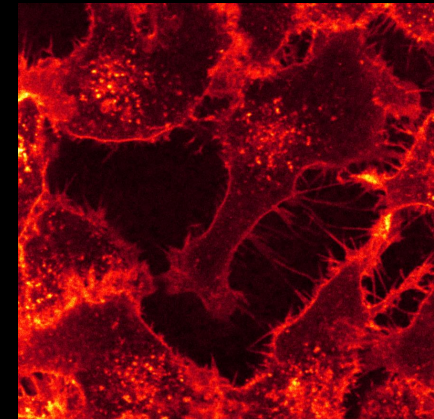
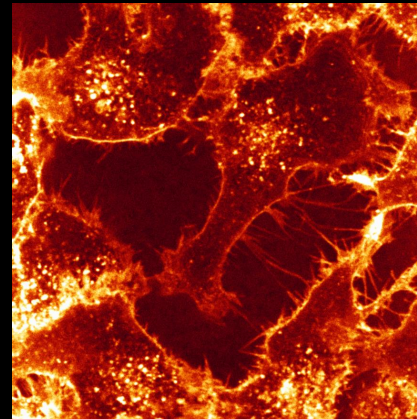
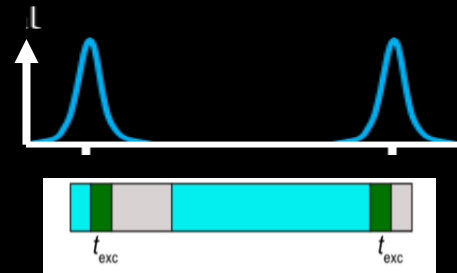
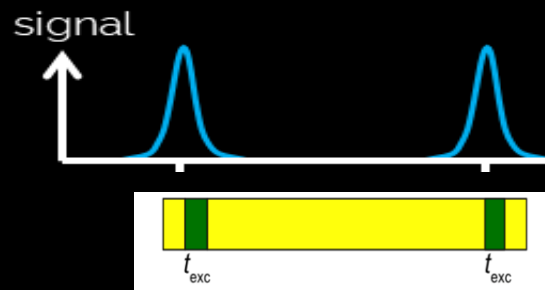
STELLARIS



The Technology Behind TauGating

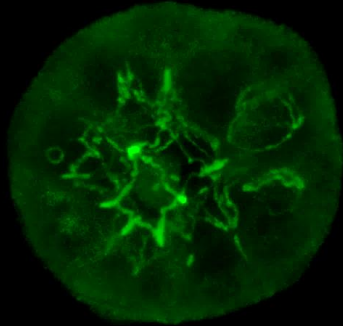


- > Digital Gate Channels (Intensity, N_{photons})

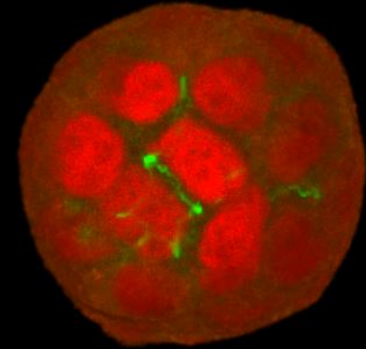
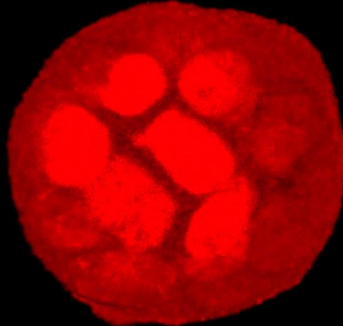


TauGating - Improve Image Quality

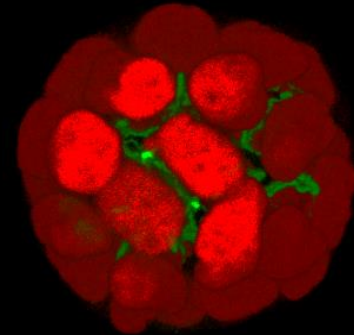
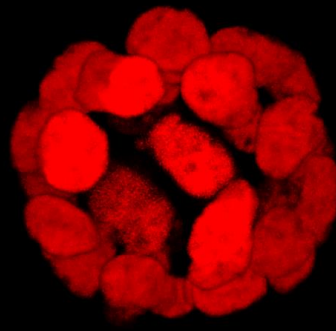
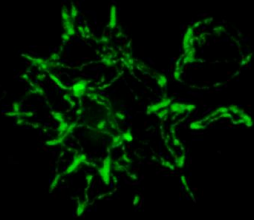
Traditional Confocal



3D cell culture

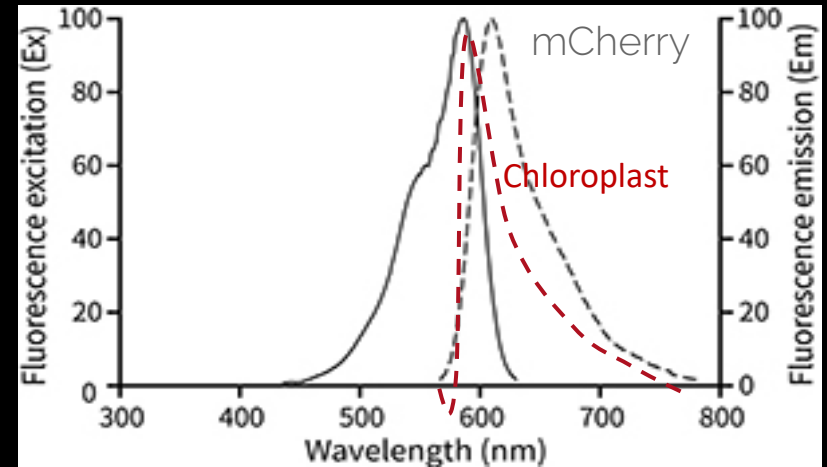
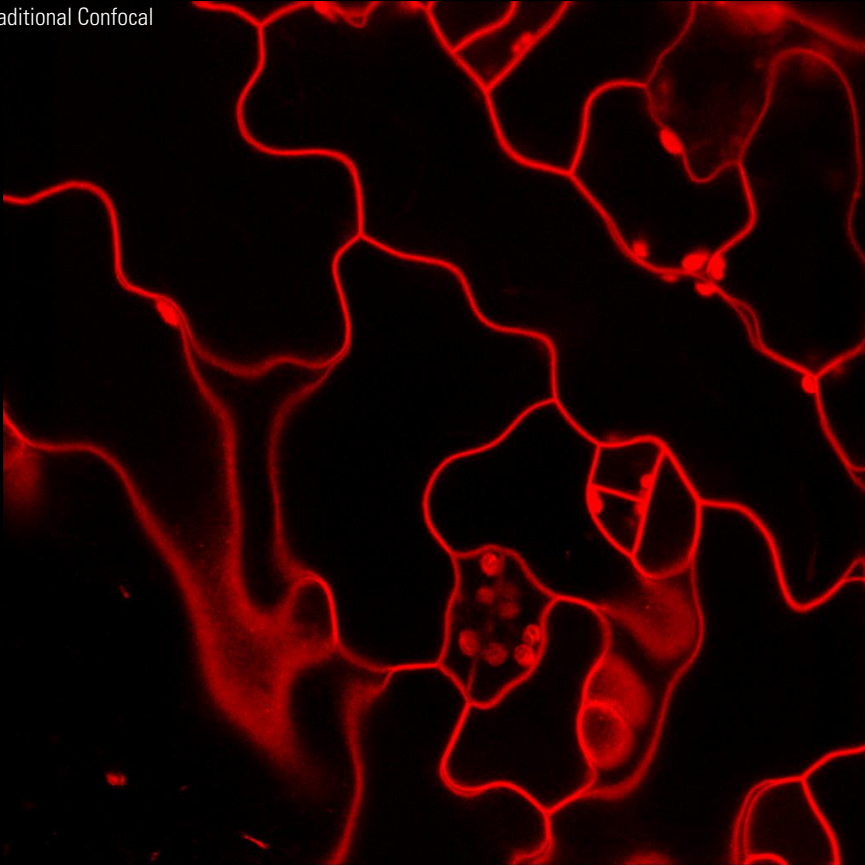


STELLARIS TauGating

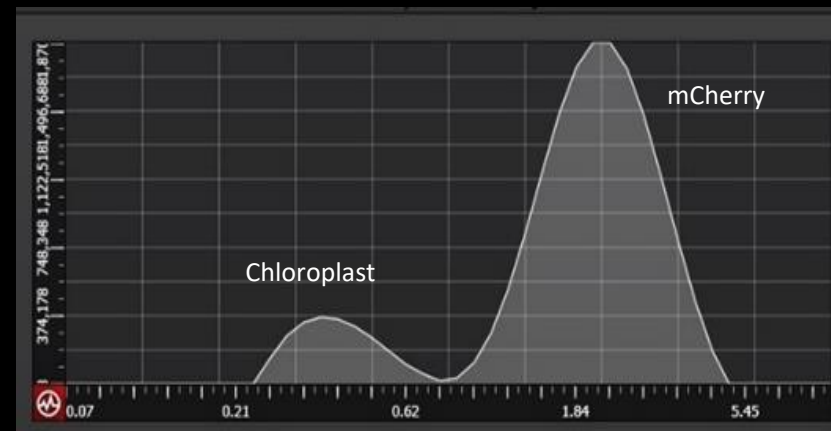


mCherry + Chloroplast

Traditional Confocal



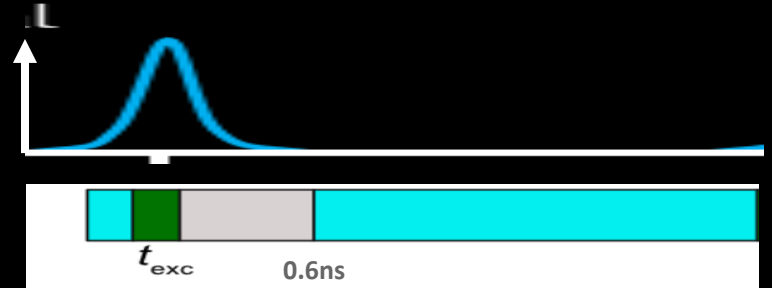
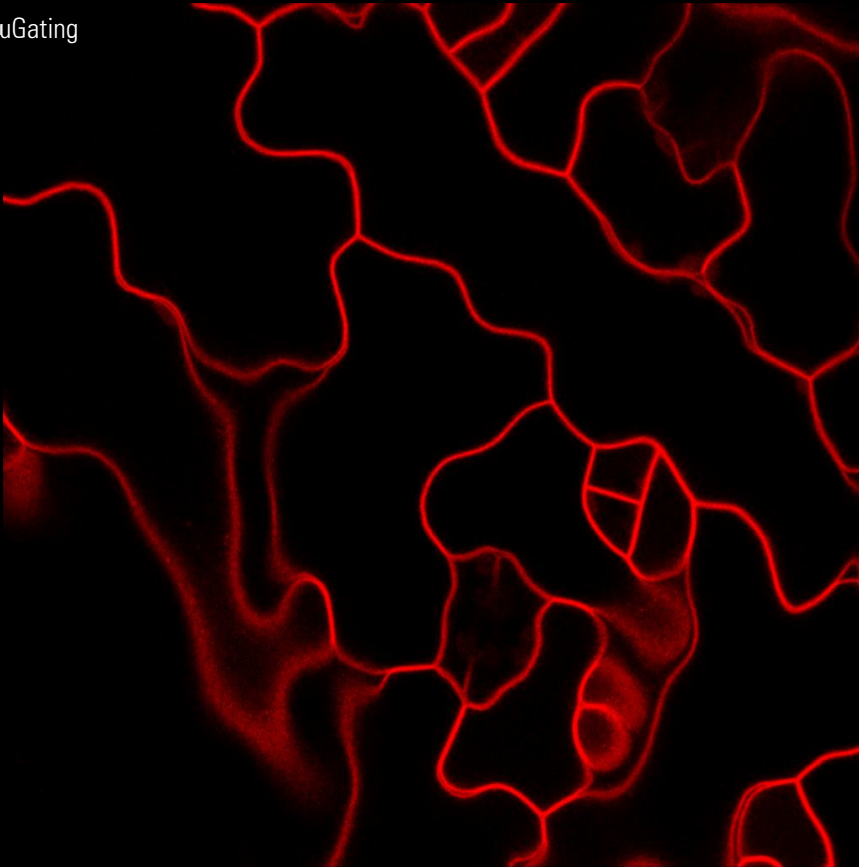
Photon counts



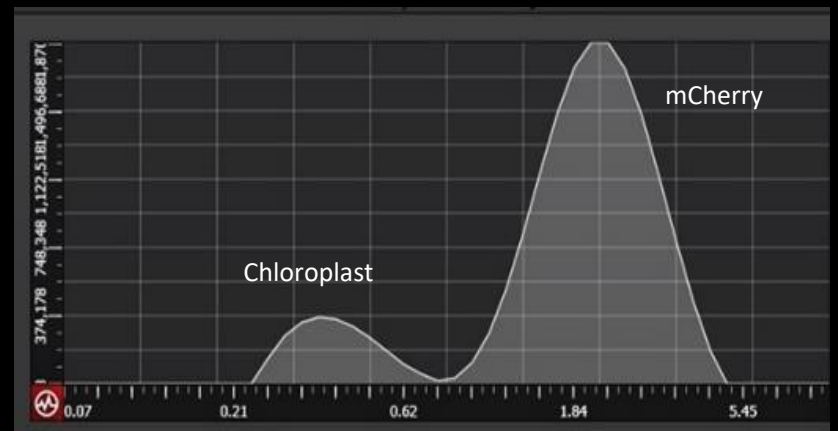
Average arrival time, ns

mCherry + Chloroplast

TauGating

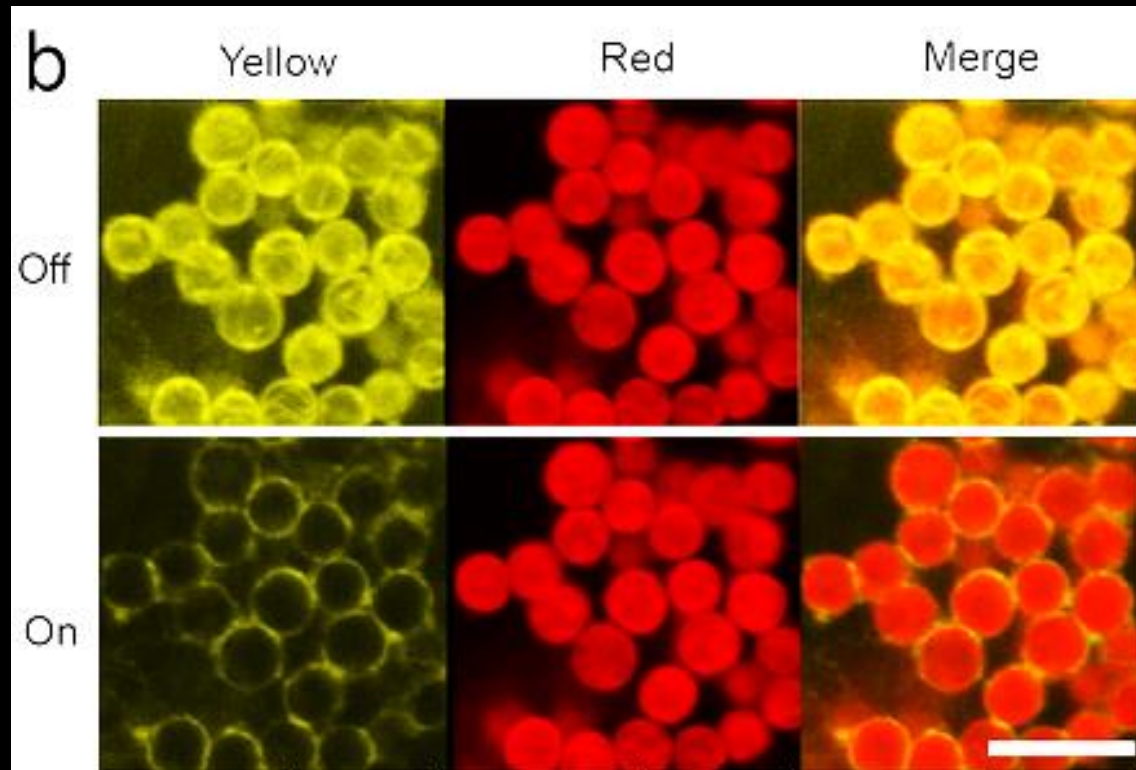


Photon counts



Average arrival time, ns

Fluorescence imaging of Mpphot-Citrine on the chloroplast periphery of transgenic *Marchantia polymorpha*.

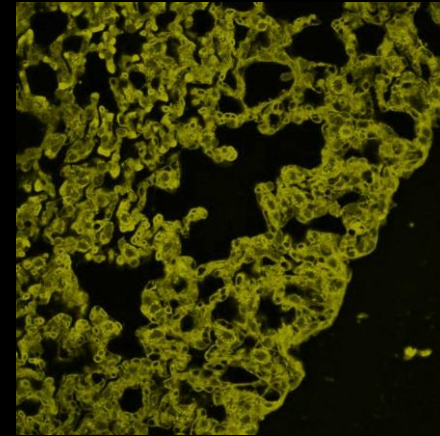
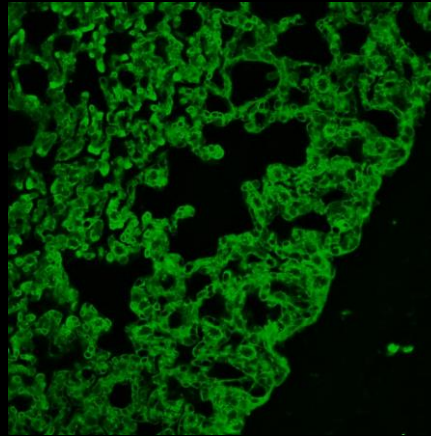
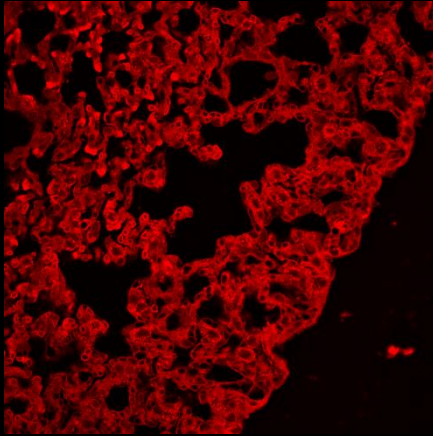


Kodama Y (2016) Time Gating of Chloroplast Autofluorescence Allows Clearer Fluorescence Imaging In Planta. PLOS ONE 11(3): e0152484. <https://doi.org/10.1371/journal.pone.0152484>
<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0152484>

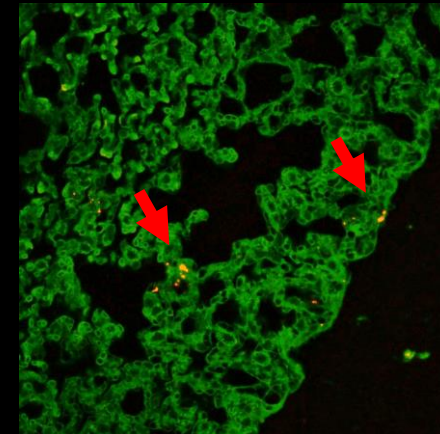
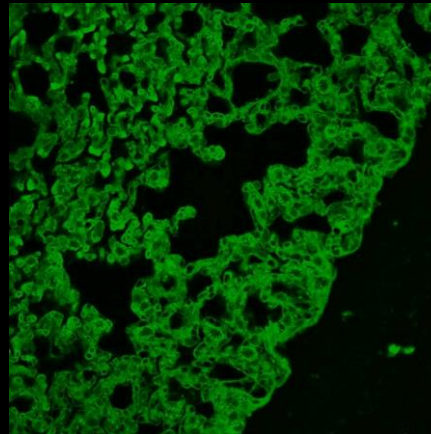
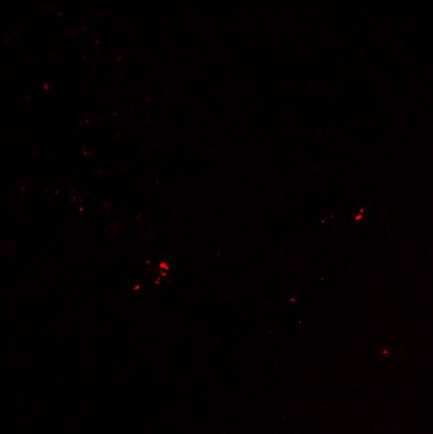
TauGating - Improve Image Quality

Tissue & nano diamond

Traditional Confocal

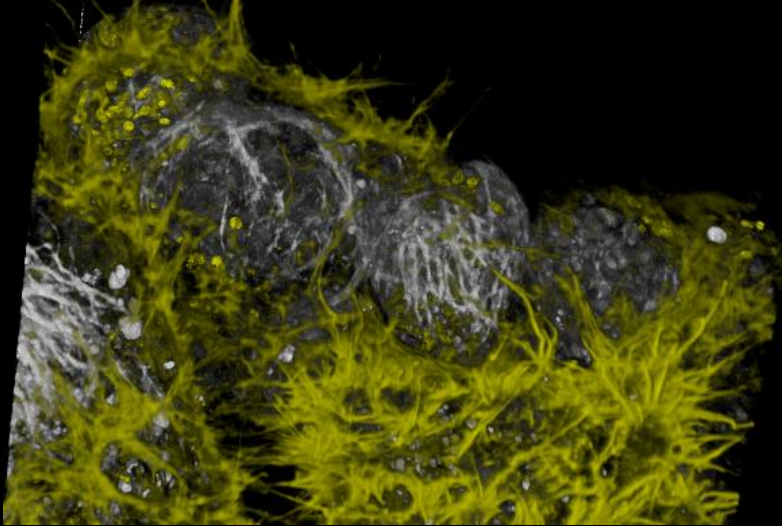


Tau gating

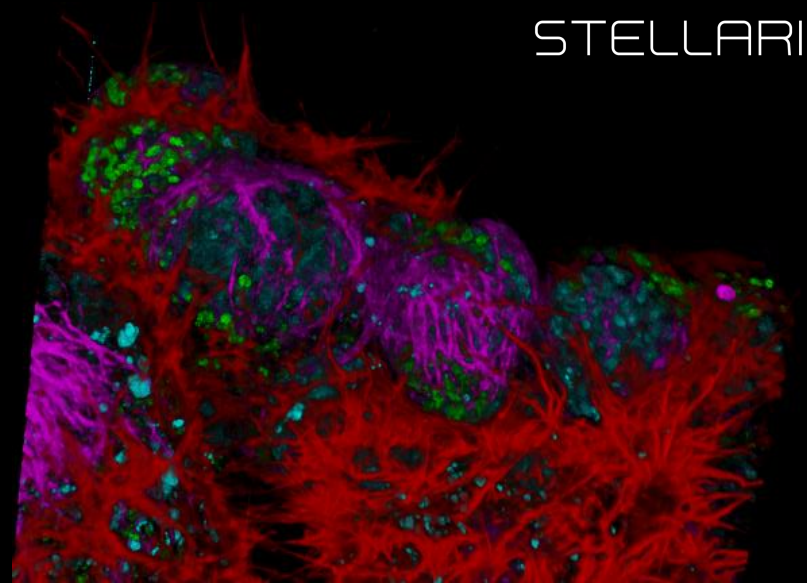


Multiplex Beyond The Spectral Options

Traditional Confocal



STELLARIS

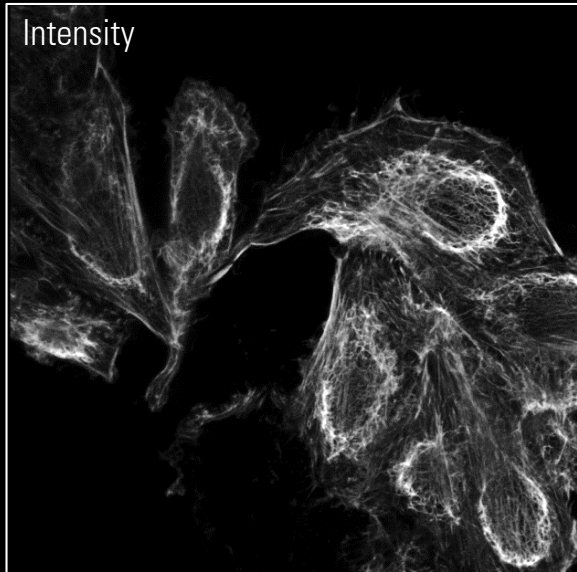


TauSense Tool:
TauSeparation

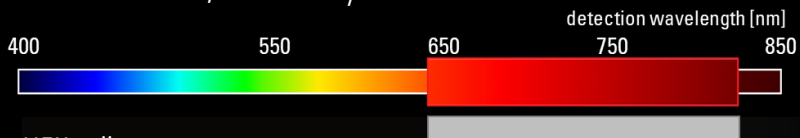


Application Example

Species Separation using TauSense



1 detector, 1 intensity channel

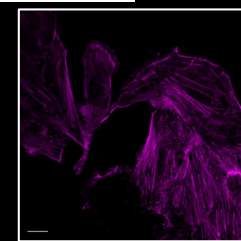
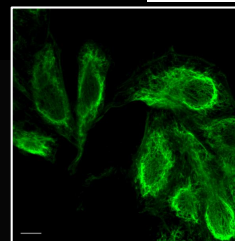
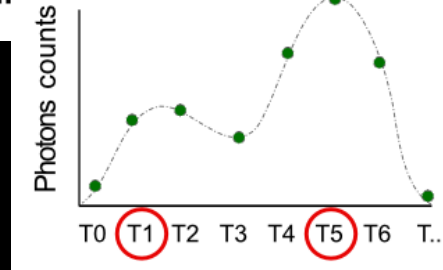
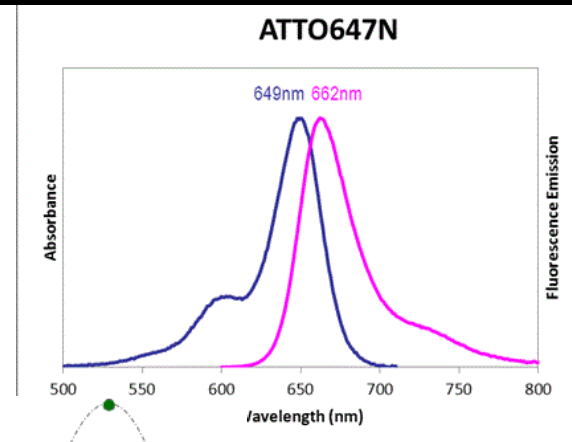
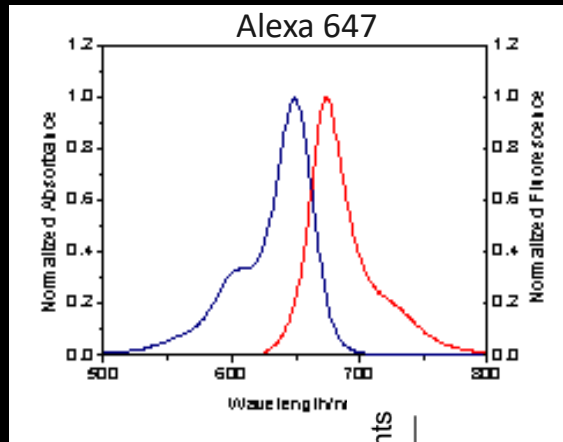


HEK cells.

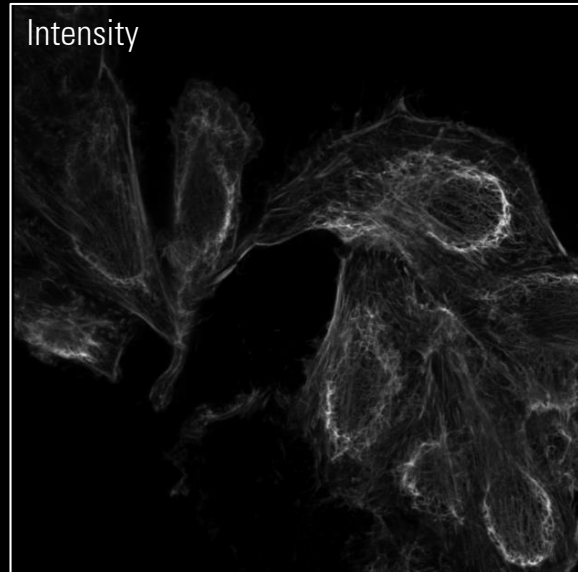
Vimentin (Alexa 647 IF), actin (ATTO647N-phalloidin)

Sample Courtesy: Sebastian Hänsch,

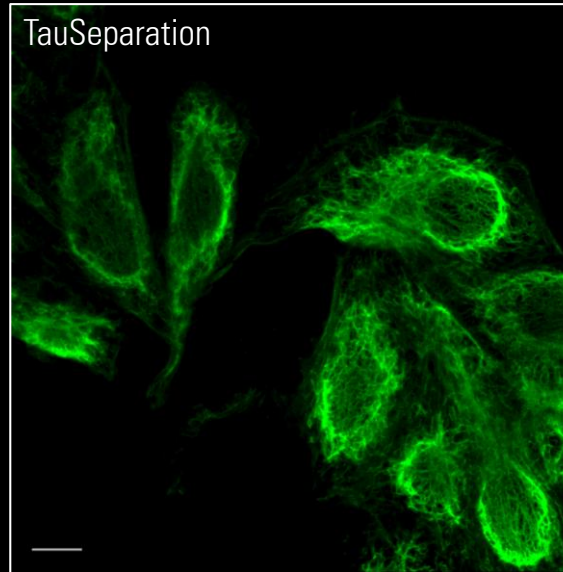
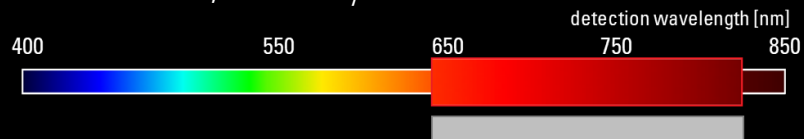
Stephanie Weidtkamp-Peters, CAI, Düsseldorf.



Species Separation using TauSense

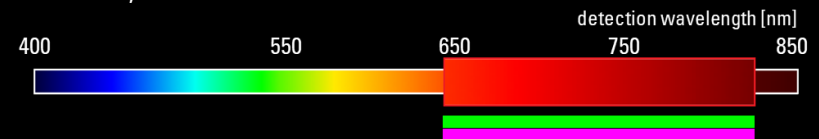


1 detector, 1 intensity channel



TauSeparation

1 detector, 2 lifetime-based channels



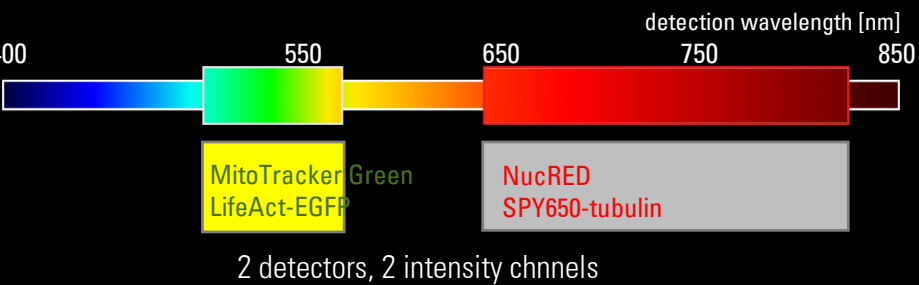
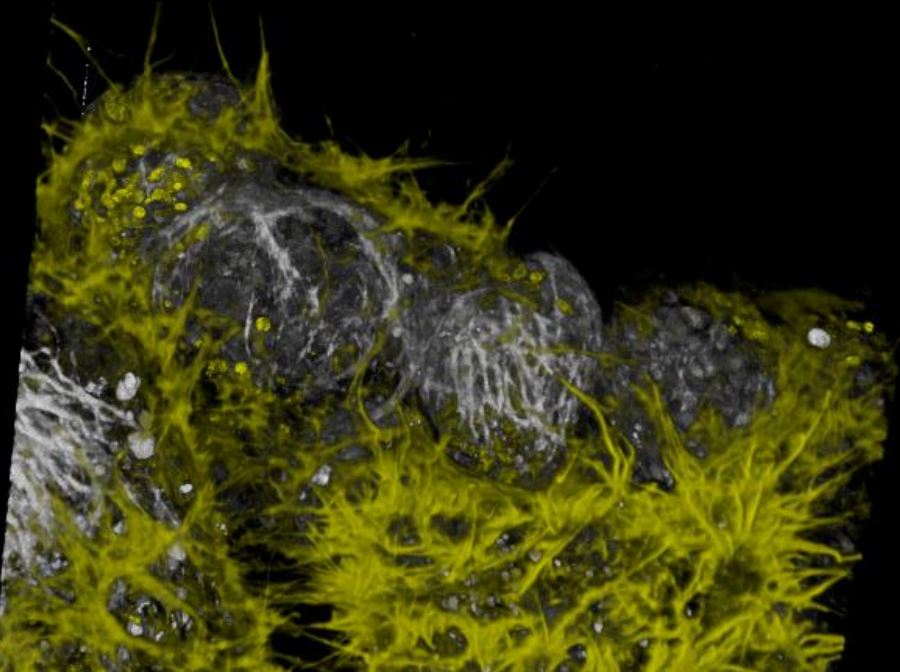
HEK cells. Vimentin (left: gray, Alexa 647 IF), actin (left: gray, ATTO647N-phalloidin). TauSeparation separates the signals coming to the detector according to the lifetime components distribution generated online at the FPGA level (right: green, Vimentin; right: magenta, Actin). Scale bar 10 μ m.

Sample Courtesy: Sebastian Hänsch, Stephanie Weidtkamp-Peters, CAI, Düsseldorf.

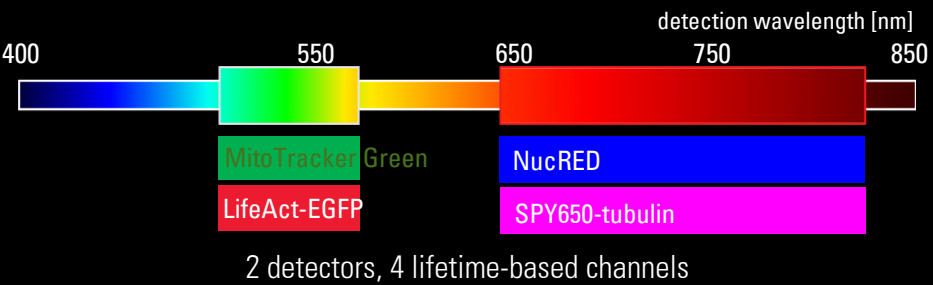
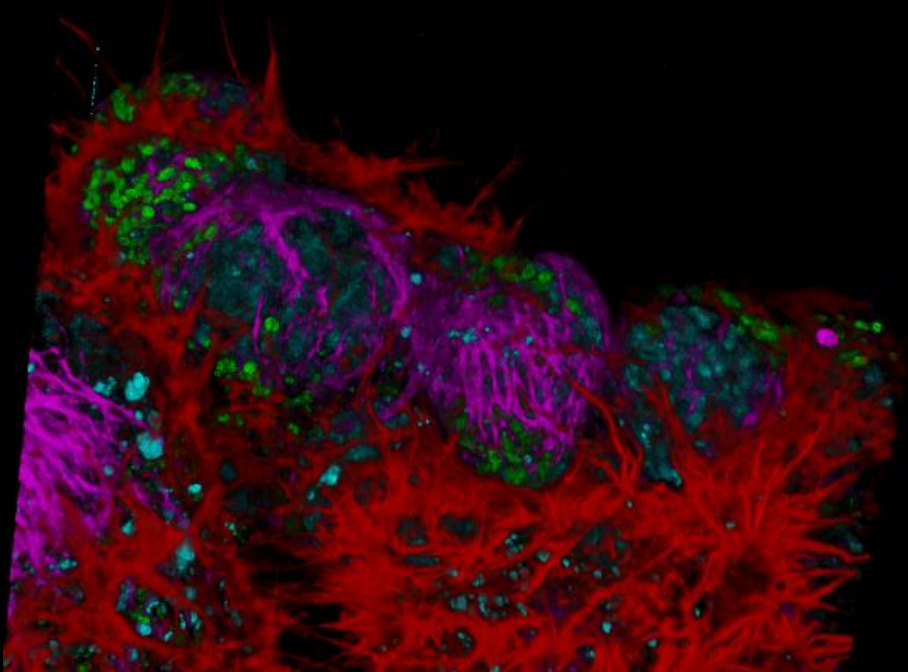


Separate Species Beyond The Spectral Options

Traditional Confocal



STELLARIS



NE-115 cells. LifeAct-mNeonGreen (left: yellow, right: red), MitoTracker Green (left: yellow, right: green), NUC Red (left: gray, right: blue), and SiR-tubulin (left: gray, right: magenta).
Courtesy: Max Heydasch, University of Bern and Spirochrome

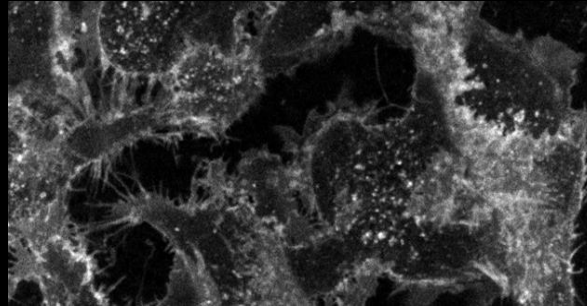


What is TauSense Good For?



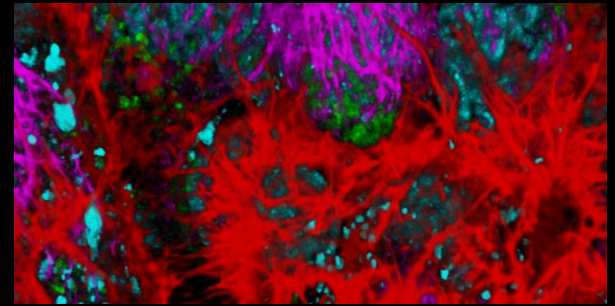
TauContrast

- Qualitative / Semi-quantitative information
- Is there a change in microenvironment? Is FRET happening?
- Changes over time (x-fold $\uparrow\downarrow$ compared to baseline)



TauGating

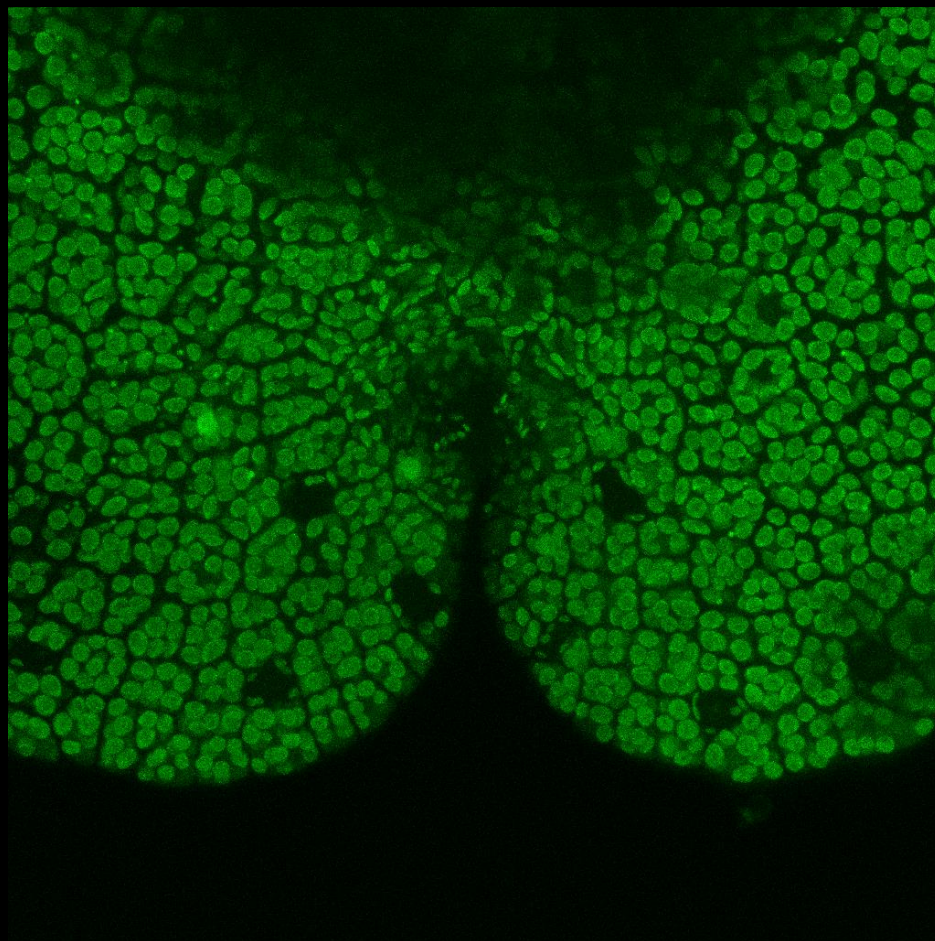
- Explore sample with gates
- Remove reflections
- Remove unwanted fluorescence contributions



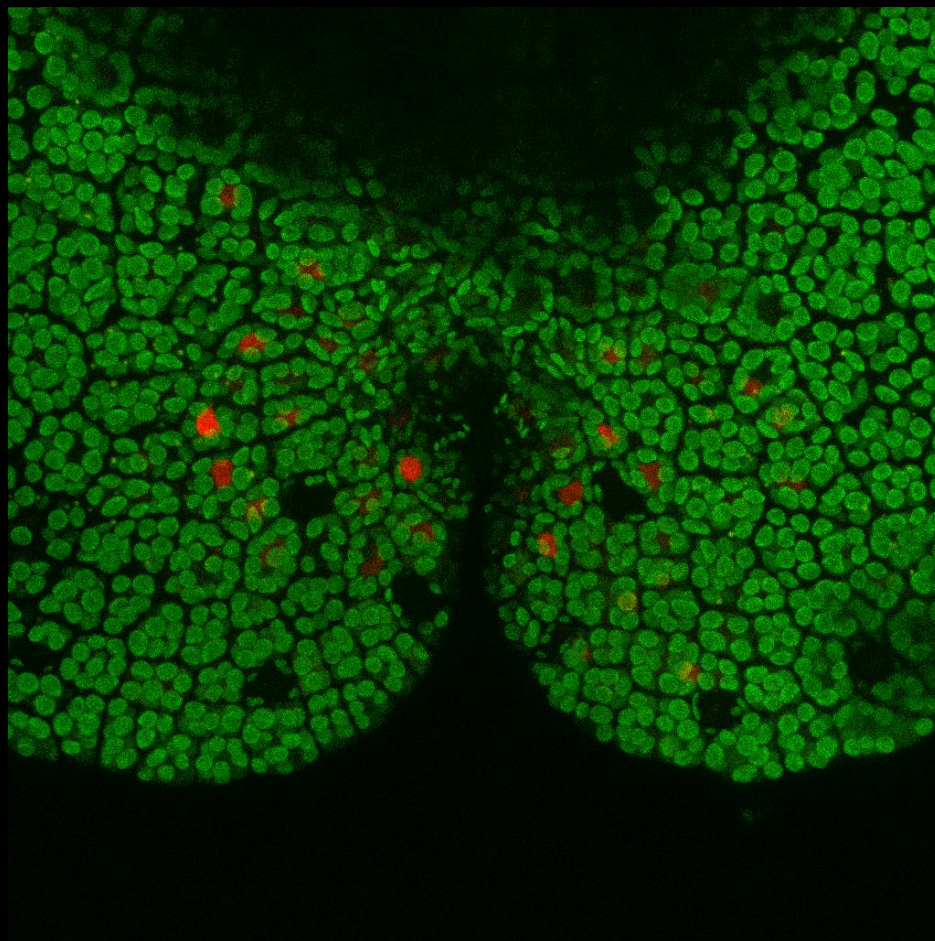
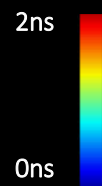
TauSeparation

- Separate species with different lifetimes

Traditional Confocal - intensity

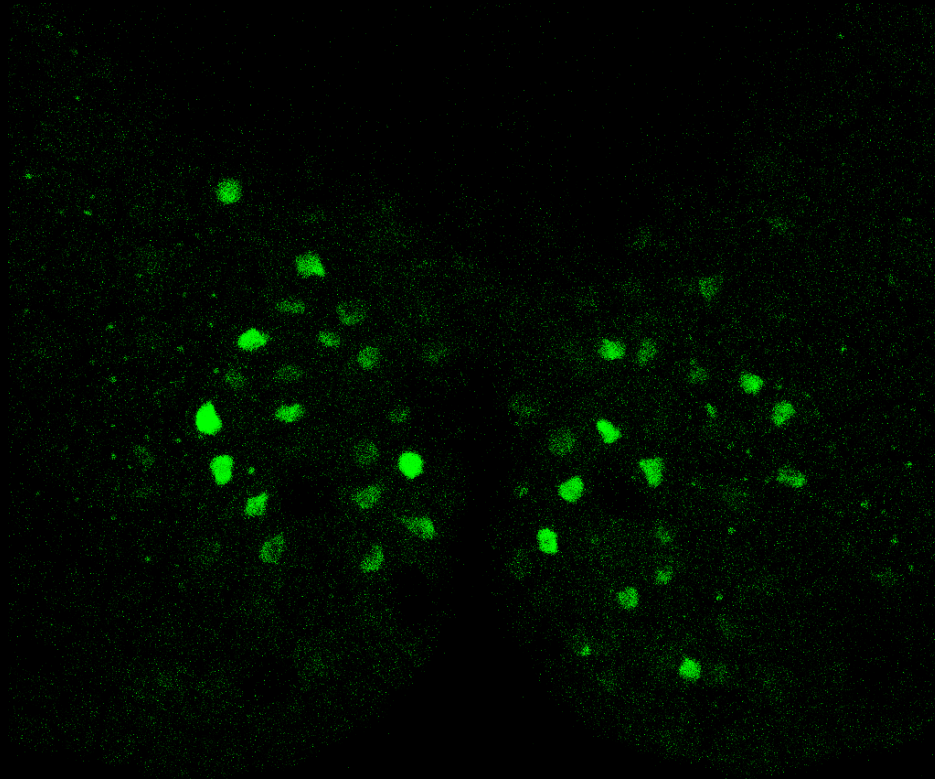


Tau Contrast

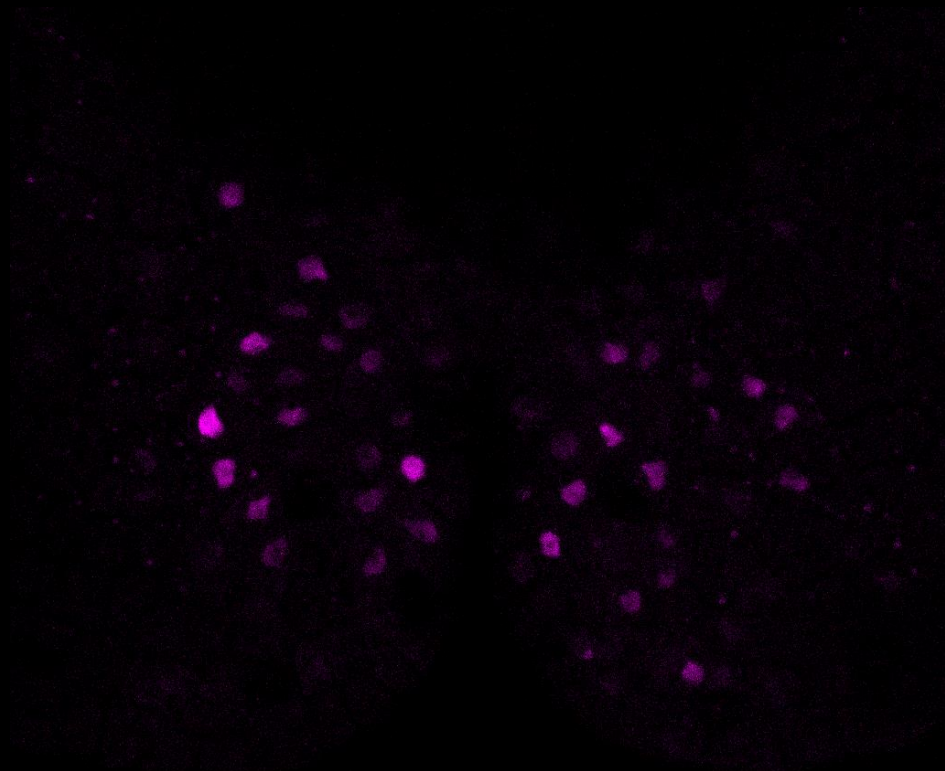
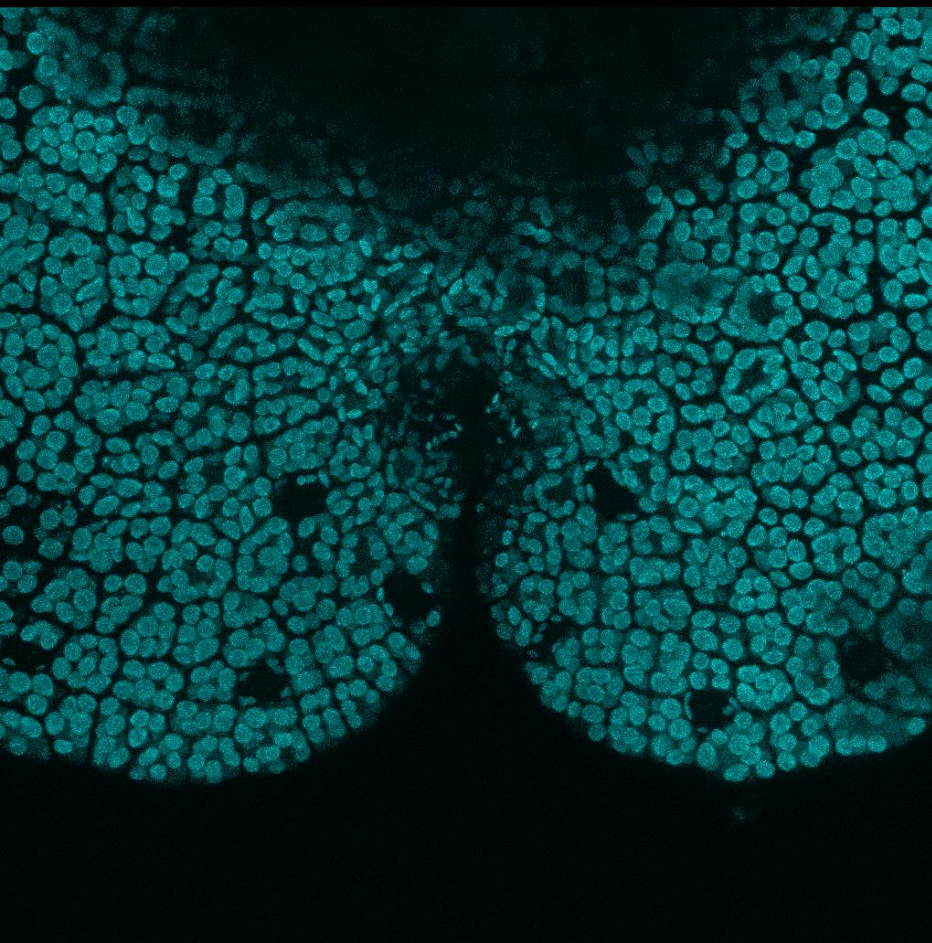


Tau Gating

Gating : 0.5-6 ns

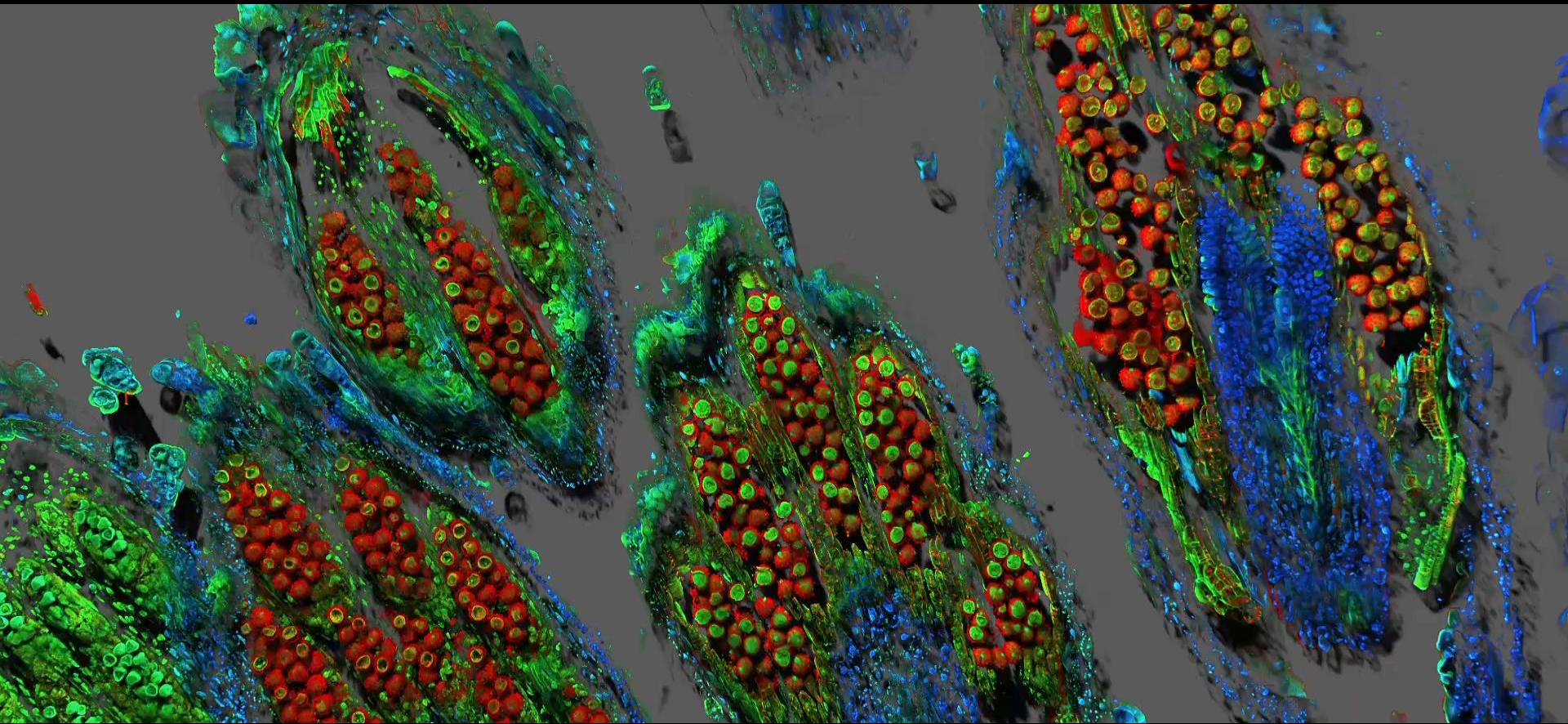


Tau Separation

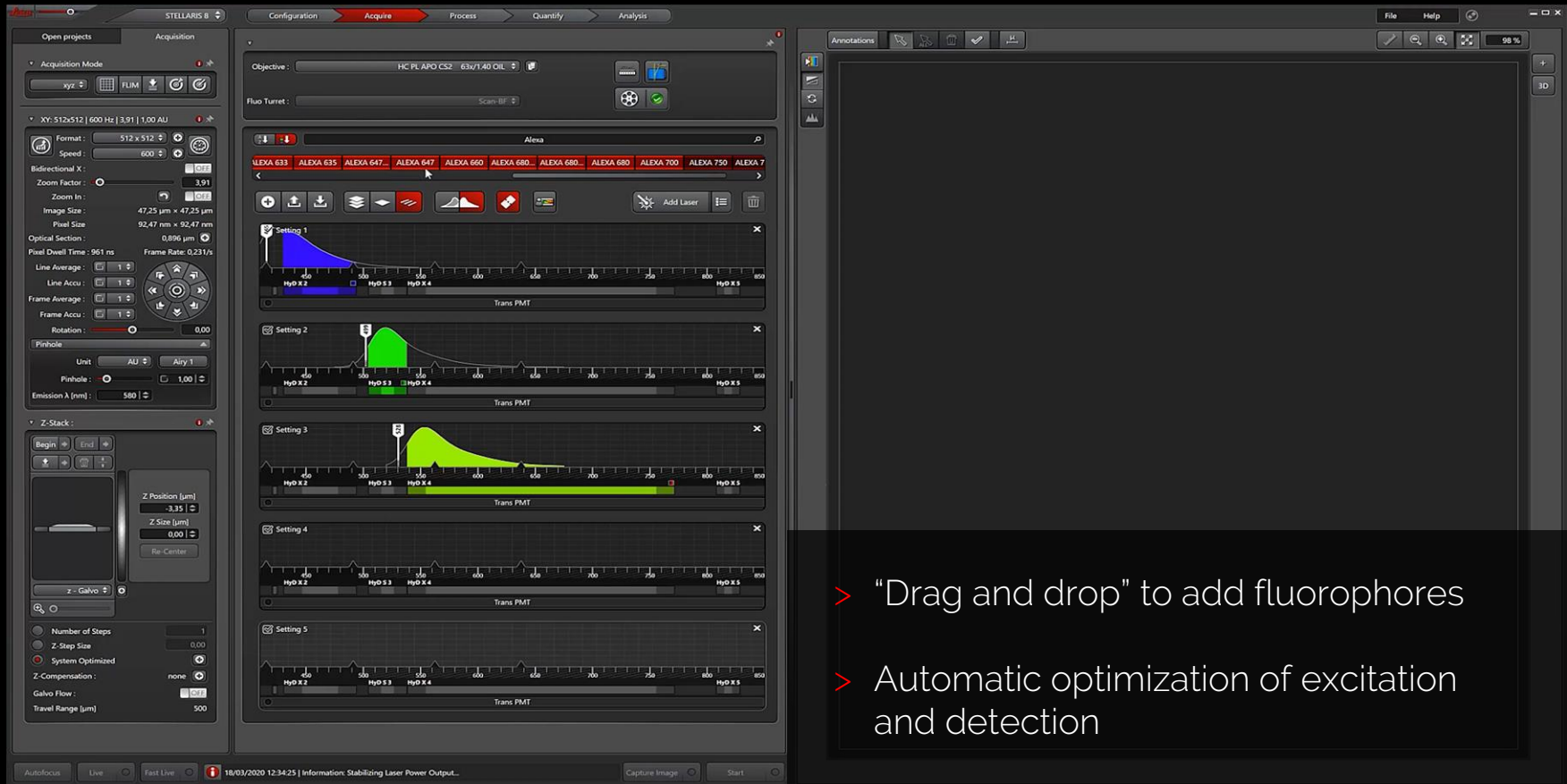


PRODUCTIVITY

DO MORE



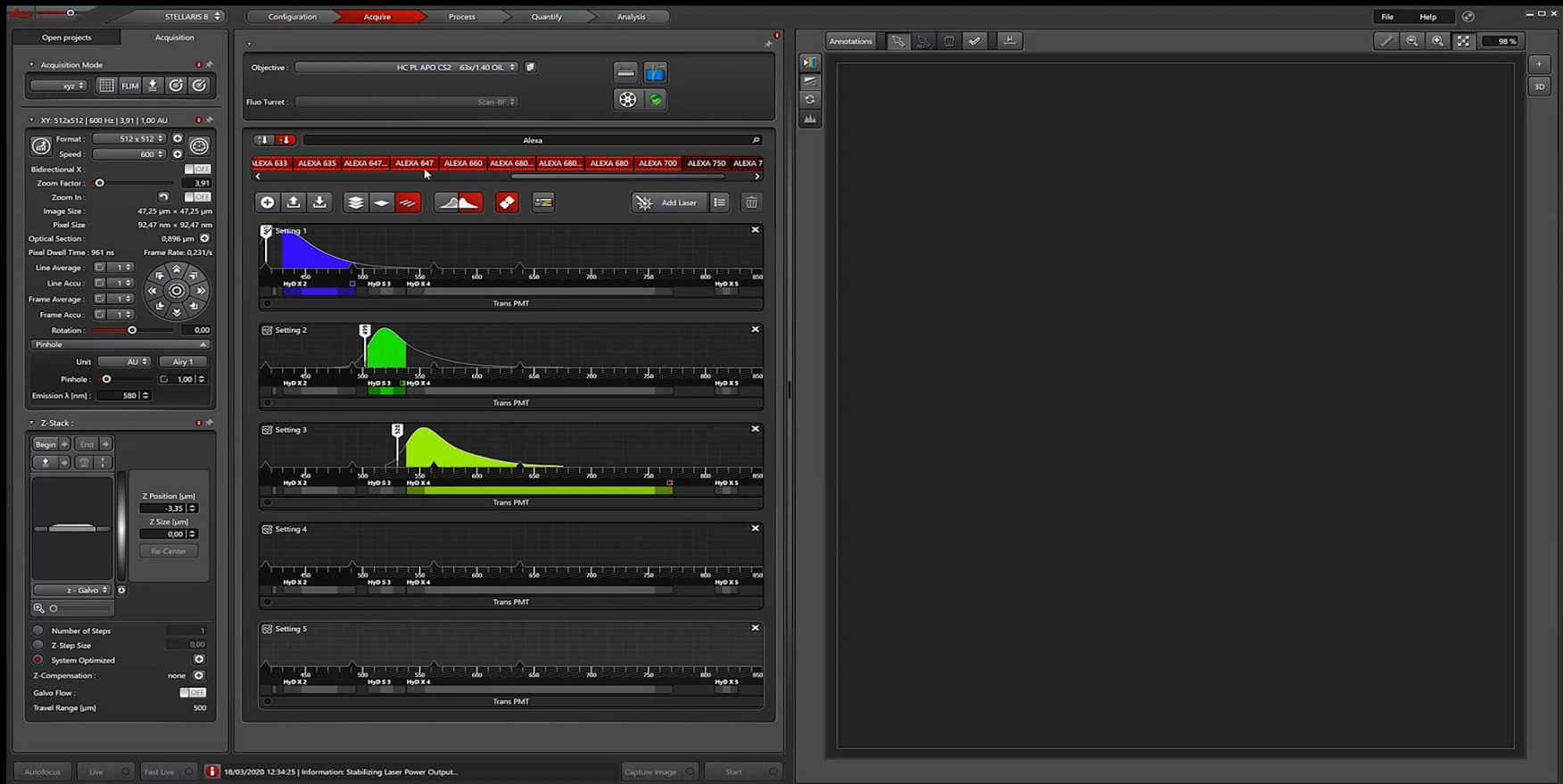
Simple, Even For Complex Experiments



The screenshot displays the Leica Stellaris 8 software interface, which is organized into several panels. On the left, the 'Acquisition' panel shows settings for 'XY: 512x512 | 600 Hz | 3.91 | 1.00 AU', 'Speed: 600', 'Zoom Factor: 3.91', and 'Image Size: 47.25 µm x 47.25 µm'. Below this, the 'Z-Stack' panel includes controls for 'Begin', 'End', 'Z Position [µm]', 'Z Size [µm]', and 'Re-Center'. The main central area is divided into two sections: the top section shows the 'Objective' (HC PLAPO CS2 63x/1.40 OIL) and 'Fluo Turret' (Scan-BF), while the bottom section displays five fluorescence spectra plots labeled 'Setting 1' through 'Setting 5'. Each plot shows intensity versus wavelength (nm) from 400 to 800 nm, with a 'Trans PMT' label. The rightmost panel contains 'Annotations' and a 'File' menu. At the bottom, a status bar indicates '18/03/2020 12:34:25 | Information: Stabilizing Laser Power Output...' and buttons for 'Capture Image' and 'Start'.

- > "Drag and drop" to add fluorophores
- > Automatic optimization of excitation and detection

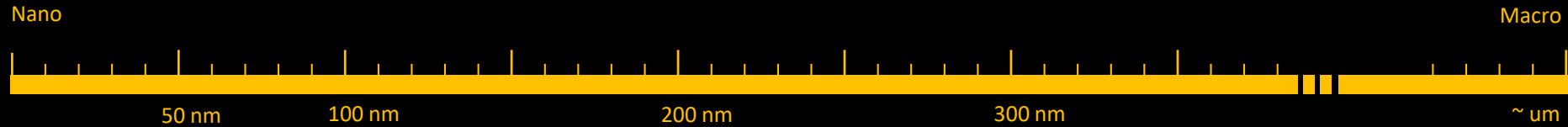
Simple, Even For Complex Experiments – Image Compass



LAS X software - Navigator



LEICA STELLARIS



30 nm

140 nm

180 nm

775STED+gSTED

Confocal SR

Confocal microscopy

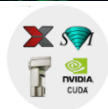
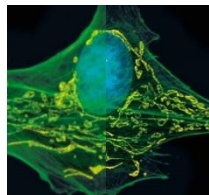
Digital light sheet

Confocal

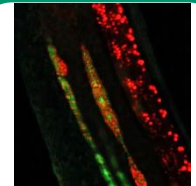
Gated STED



**775 pulsed
STED**



Lightning



DLS

STELLARIS

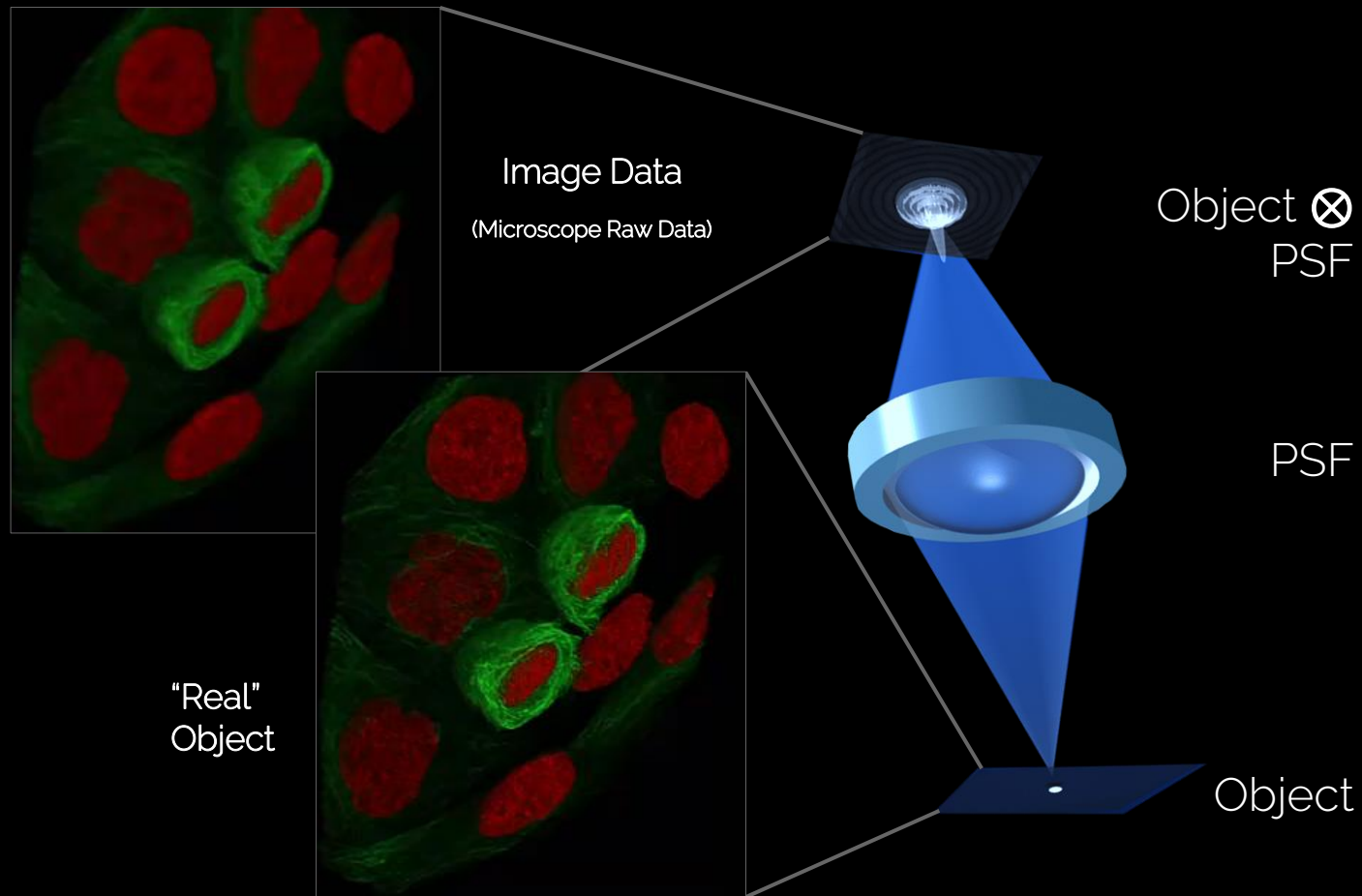


LIGHTNING

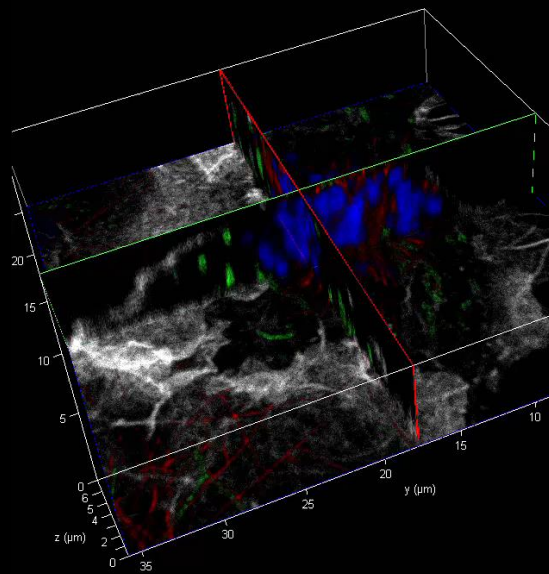
Image Information Extraction



Microscopy Image Formation

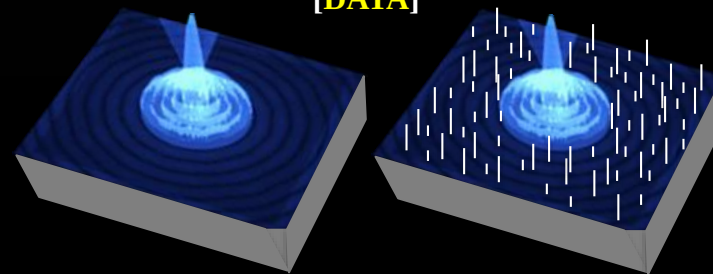
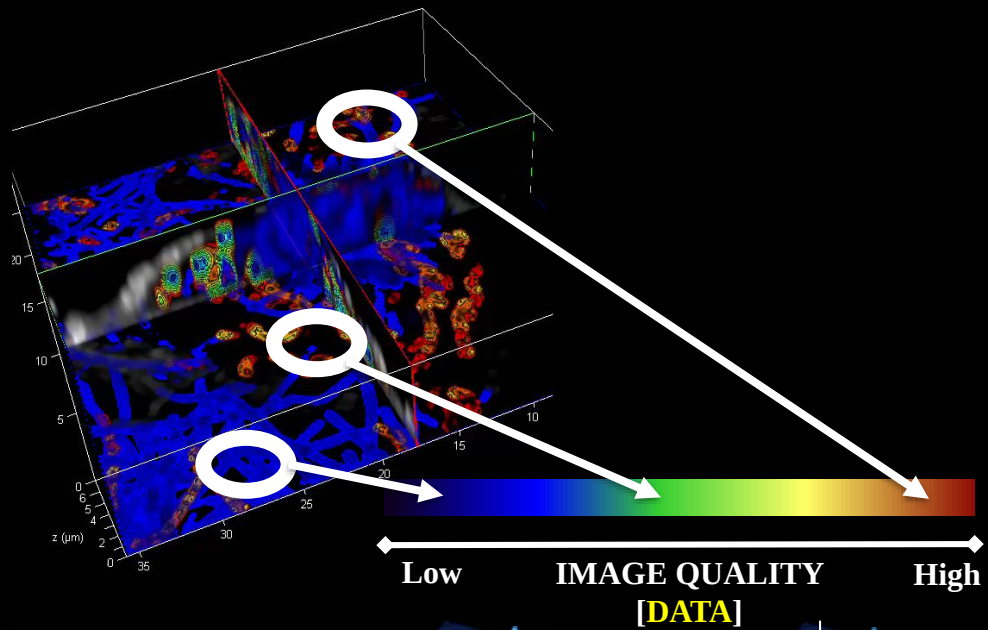


Raw data

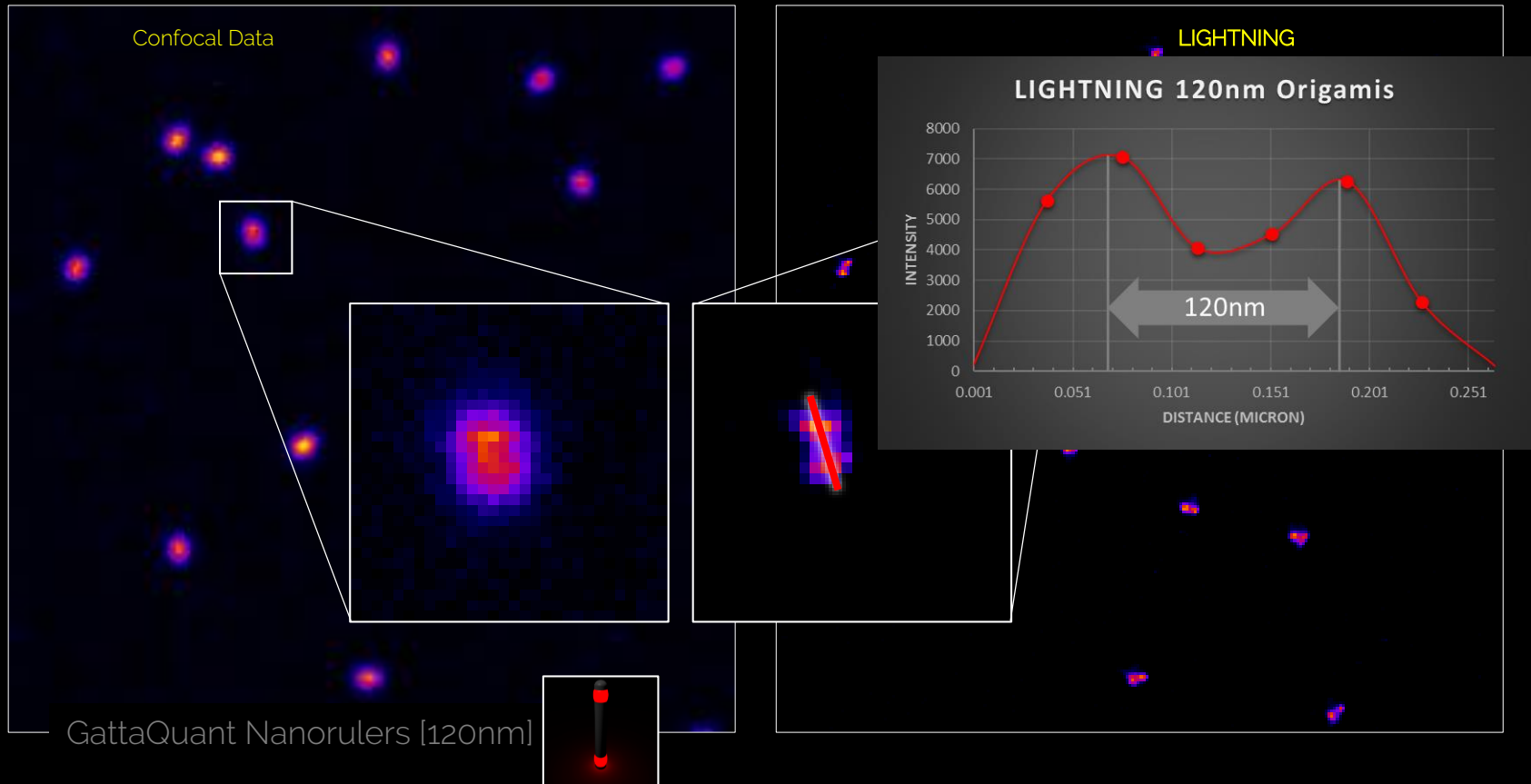


GATTA cells

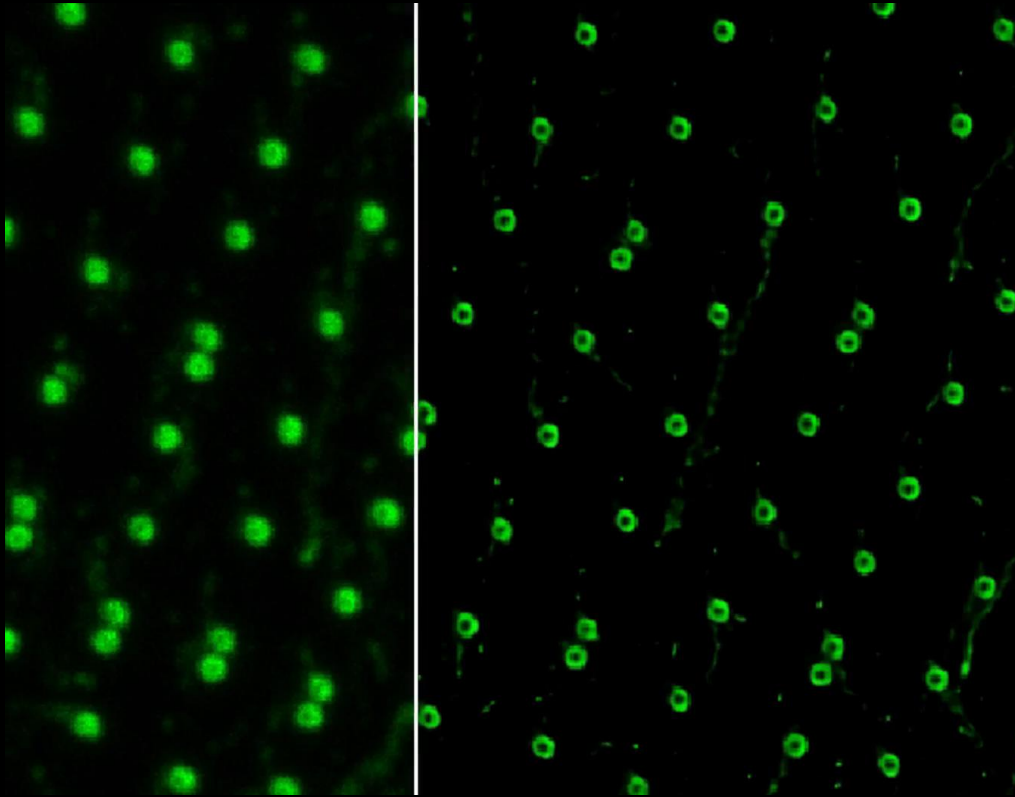
Decision Mask
Position dependant image quality



Accessing Super-Resolution



LIGHTNING: Accessing The True Nature Of Image Data



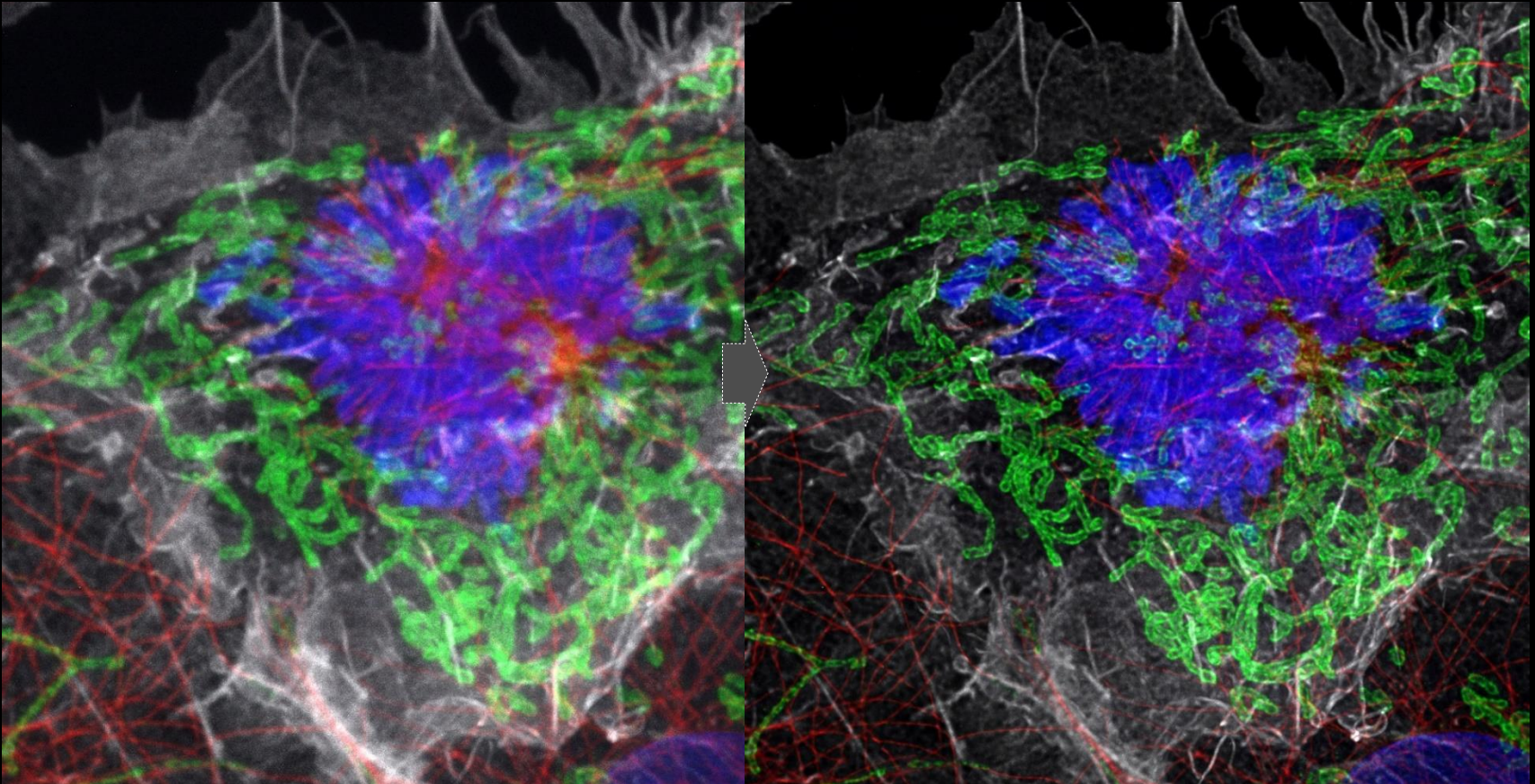
Confocal | MP | gated STED

Including every imaging modality



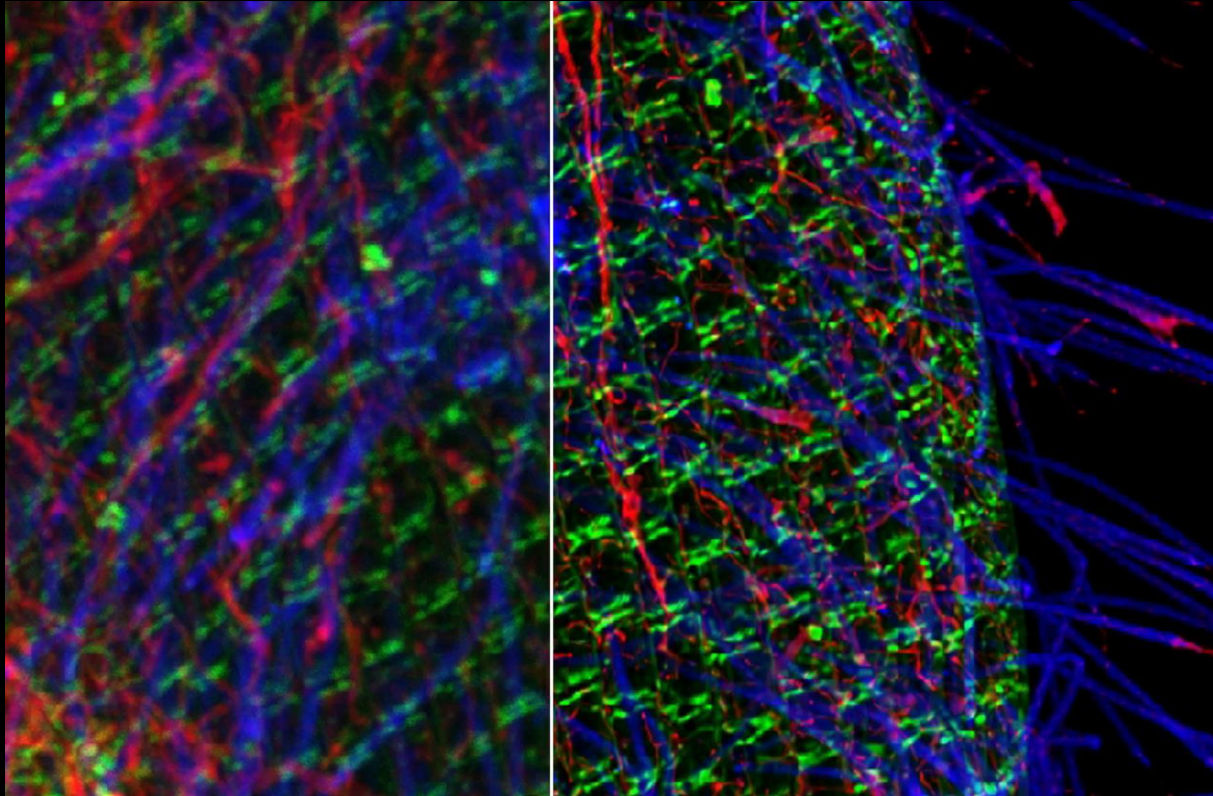
LIGHTNING: Adaptive Multicolor Super-Resolution

Adaptive Deconvolution



Gatta cells

LIGHTNING: Adaptive Multicolor Super-Resolution



Confocal | MP | gated STED

Including every imaging modality



Key Features of Lightning

-Leica Confocal Superresolution Microscopy-

- $\text{dxy} \sim 120\text{nm}$ vs dxy confocal $\sim 180\text{nm}$;
 $\text{dz} \sim 350\text{nm}$ vs dz confocal $\sim \lambda (500\text{nm})$
4x contrast better
- **Simultaneously multicolor confocal superresolution**
- **As same speed as confocal scan**
- High-speed multicolor live imaging
- Std. application for STELLARIS

STELLARIS

